

Regulation of Epigenomes

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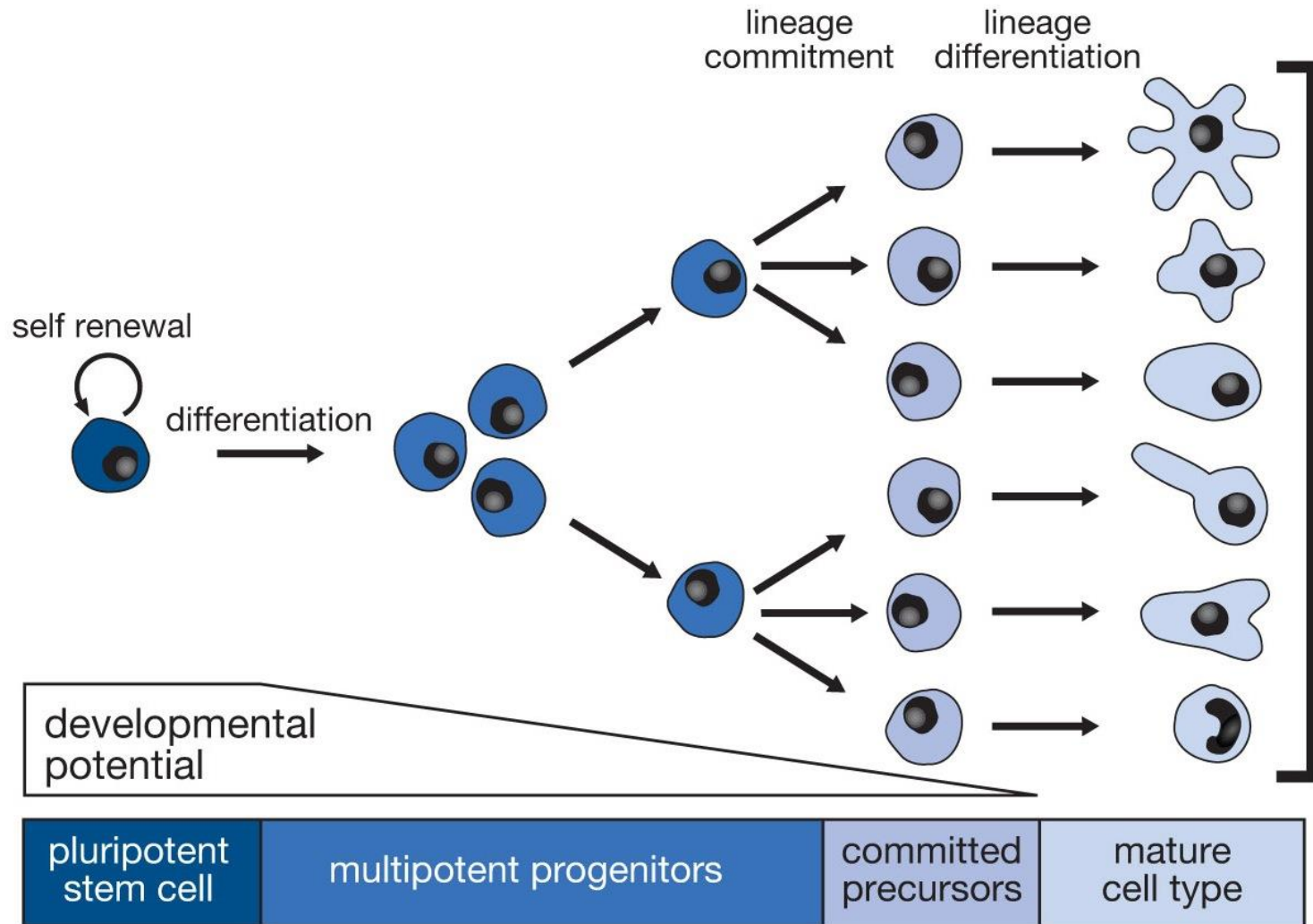
Summary

- **Classical examples of epigenetic regulations**
- **Chromatin**
- **Epigenetic inheritance**
- **Epigenetic changes in cancer**
- **Methods to detect epigenetic changes**

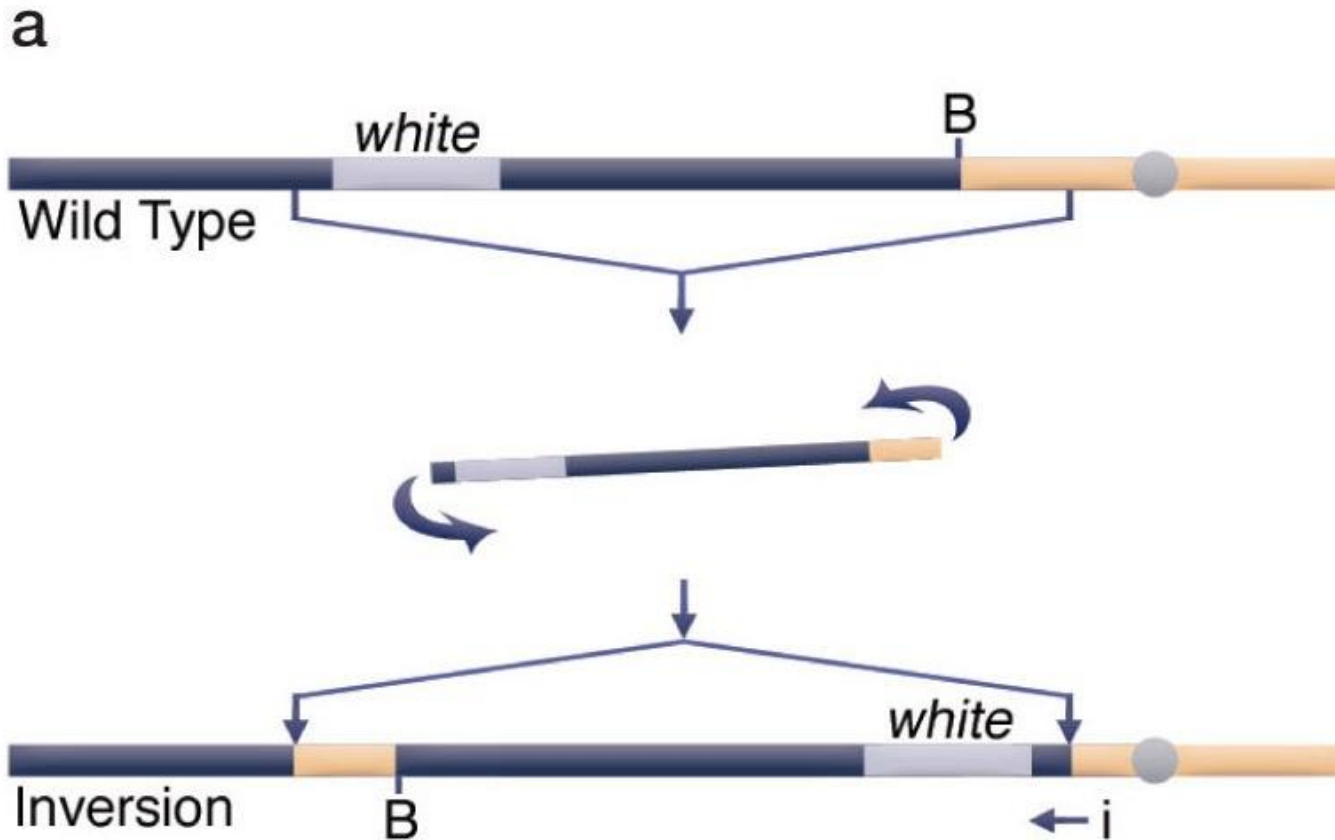
Epigenetics

- **Coined by Waddington in 1942 to explain differentiation of cells from one state to another**
- **Epigenetics: heritable changes in gene expression without changes in DNA sequences.**

Epigenetic regulation of gene expression during development

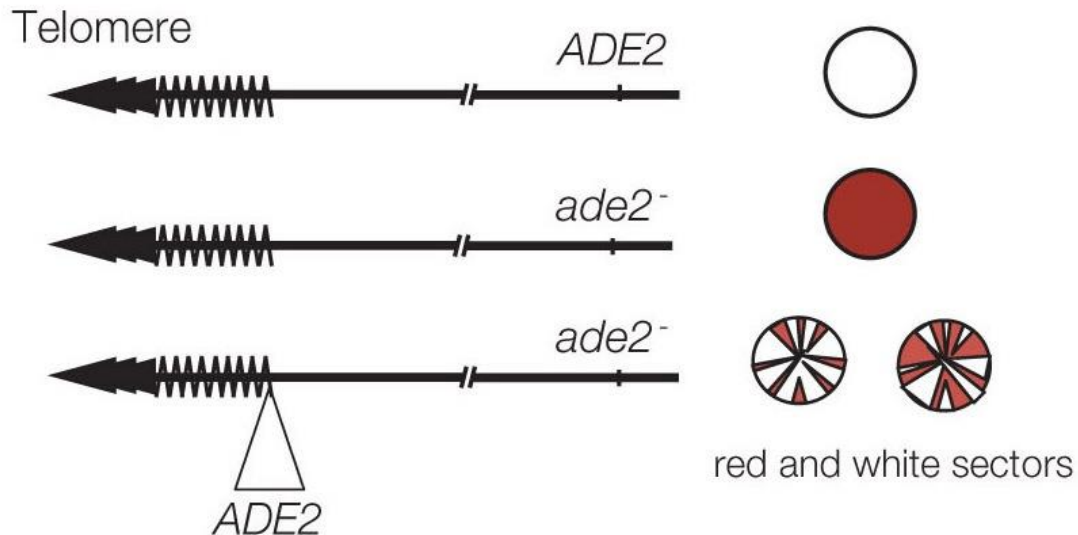


Position effect variegation in *Drosophila*

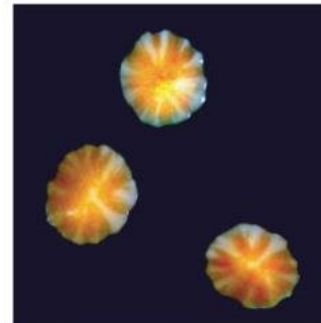


Telomere position effect (TPE): a form of epigenetic silencing

TPE of *ADE2* expression in *S.cerevisiae*



ade2⁻; ADE2-TelVR
variegated
repression



Colors of flowers, animal furs.....



Picotee



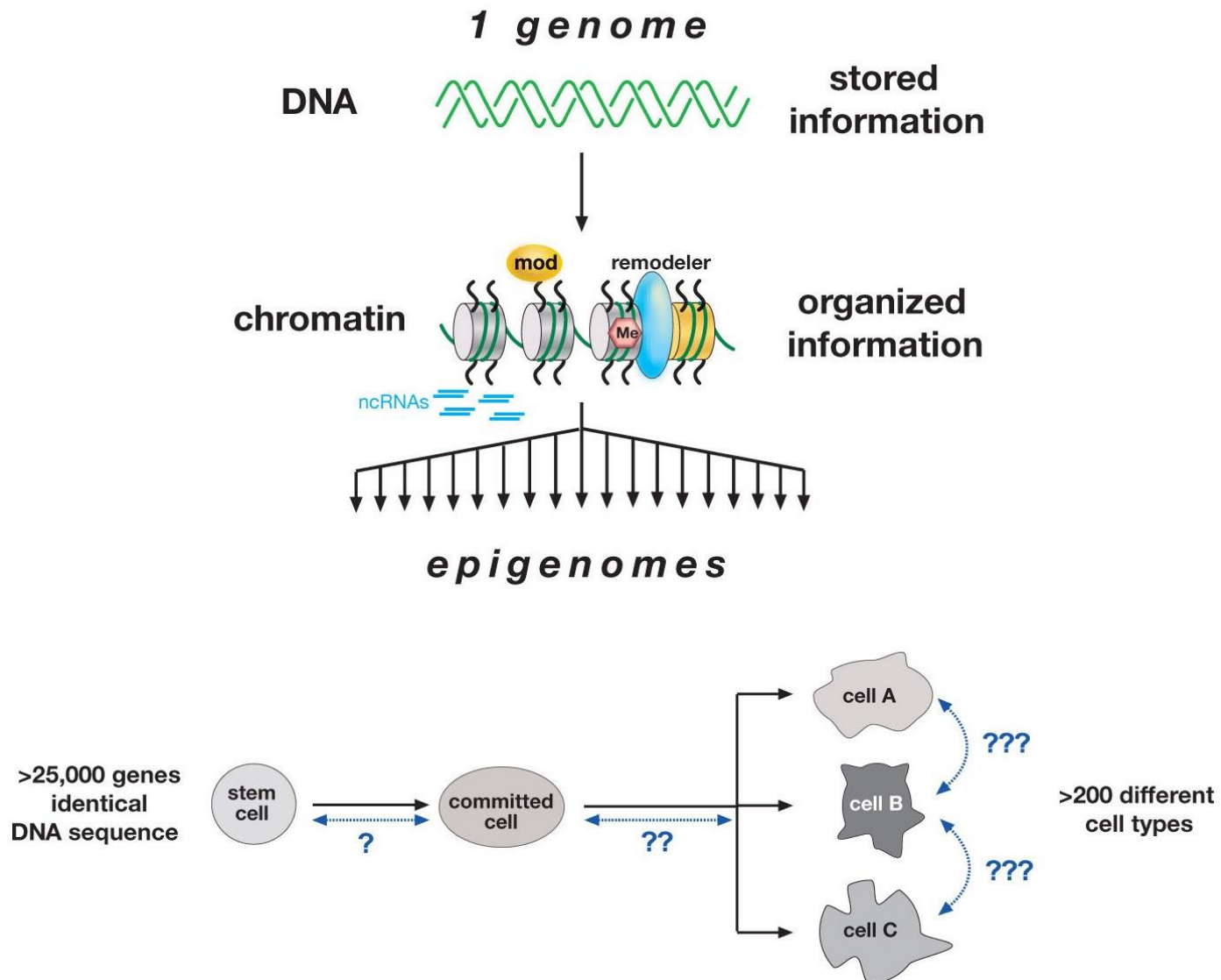
Star



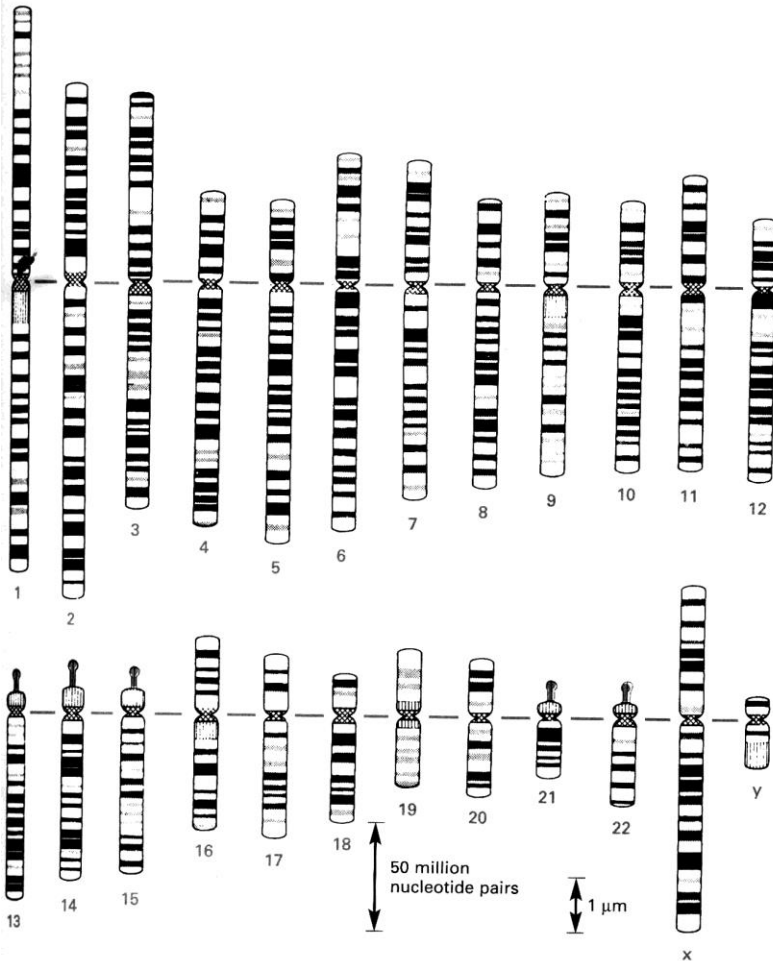
'Night Sky'

Morita and Hoshino, Breeding Science, 2018

One genome and many epigenomes



Chromatin encodes epigenetic information



- Chromatin is a complex of DNA and proteins
- Genetic information refers to DNA sequences
- Epigenetic information is “stored” in and regulated by chromatin structures

Euchromatin and heterochromatin



Euchromatin

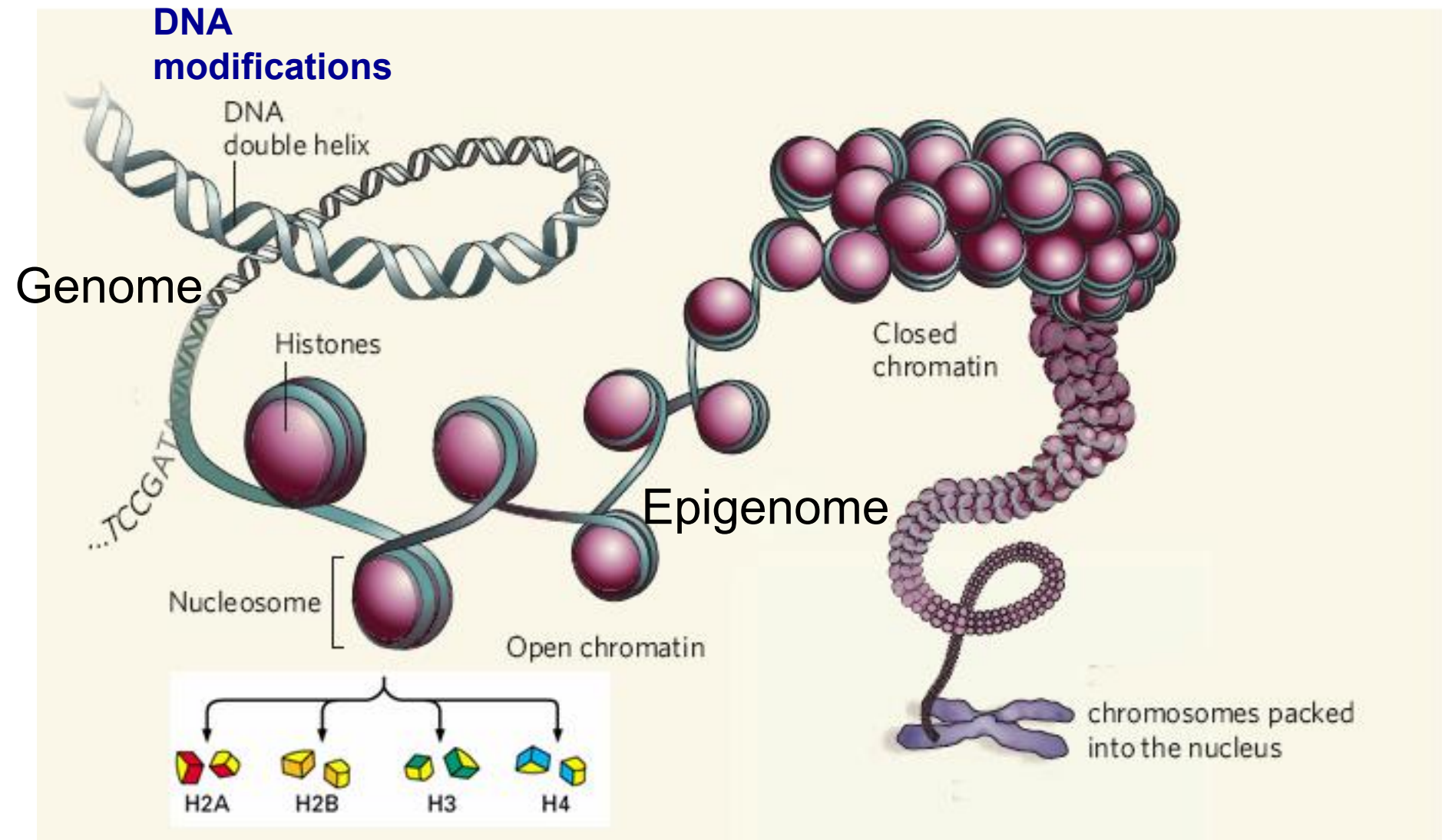
- Regions “rich” in genes
- Regions active in transcription
- Hyperacetylated histones
- Replicate early in S phase of the cell cycle

Heterochromatin

- Regions “poor” in genes
- Regions that silence transcription
- Hypoacetylated histones
- Replicate late S phase

At molecular levels, chromatin domains can be classified based on modifications on histones as well as chromatin binding proteins

DNA is packaged into chromatin in eukaryotes

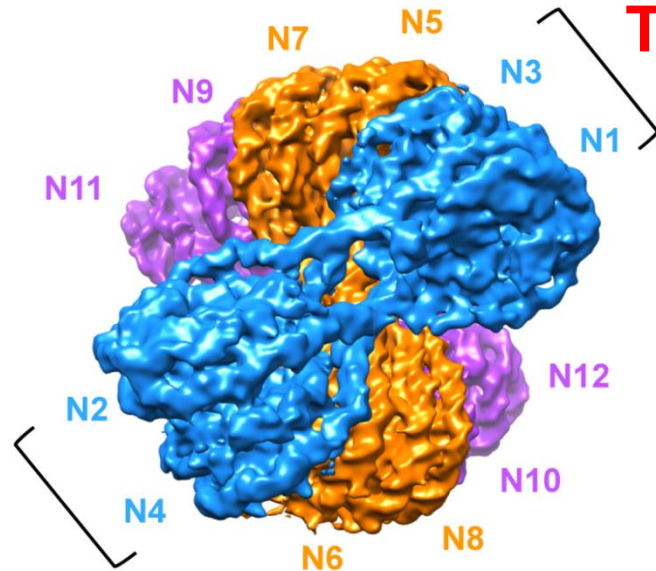


Nucleosome

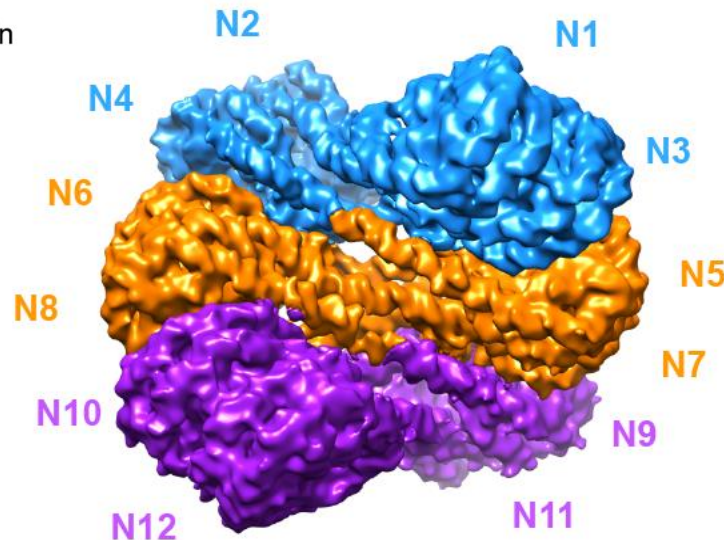
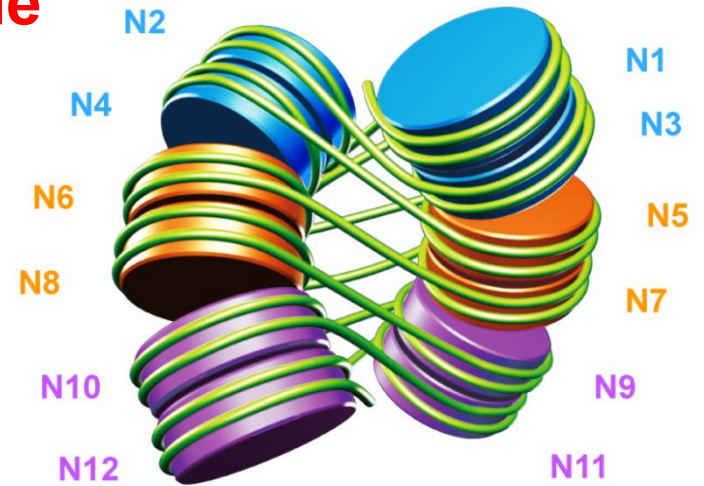


“Tetranucleosome” is the structural unit of chromatin fiber

Tetranucleosome



12×187 bp chromatin

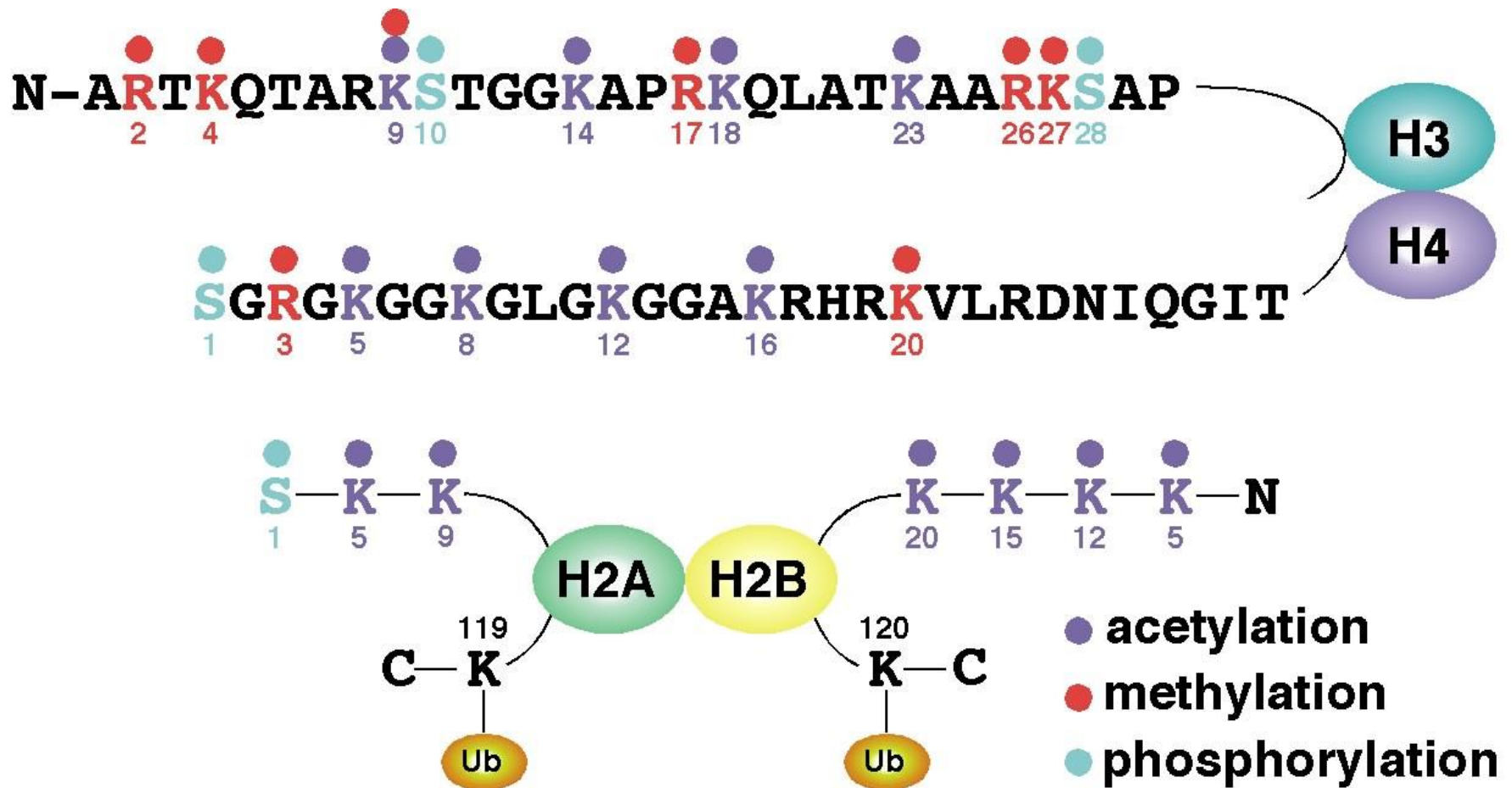


**Left-handed
twist of
tetranucleosomal
unit**

Epigenetic Marks

- DNA methylation, hydroxymethylation
- Histone modifications
- Non-coding RNAs?

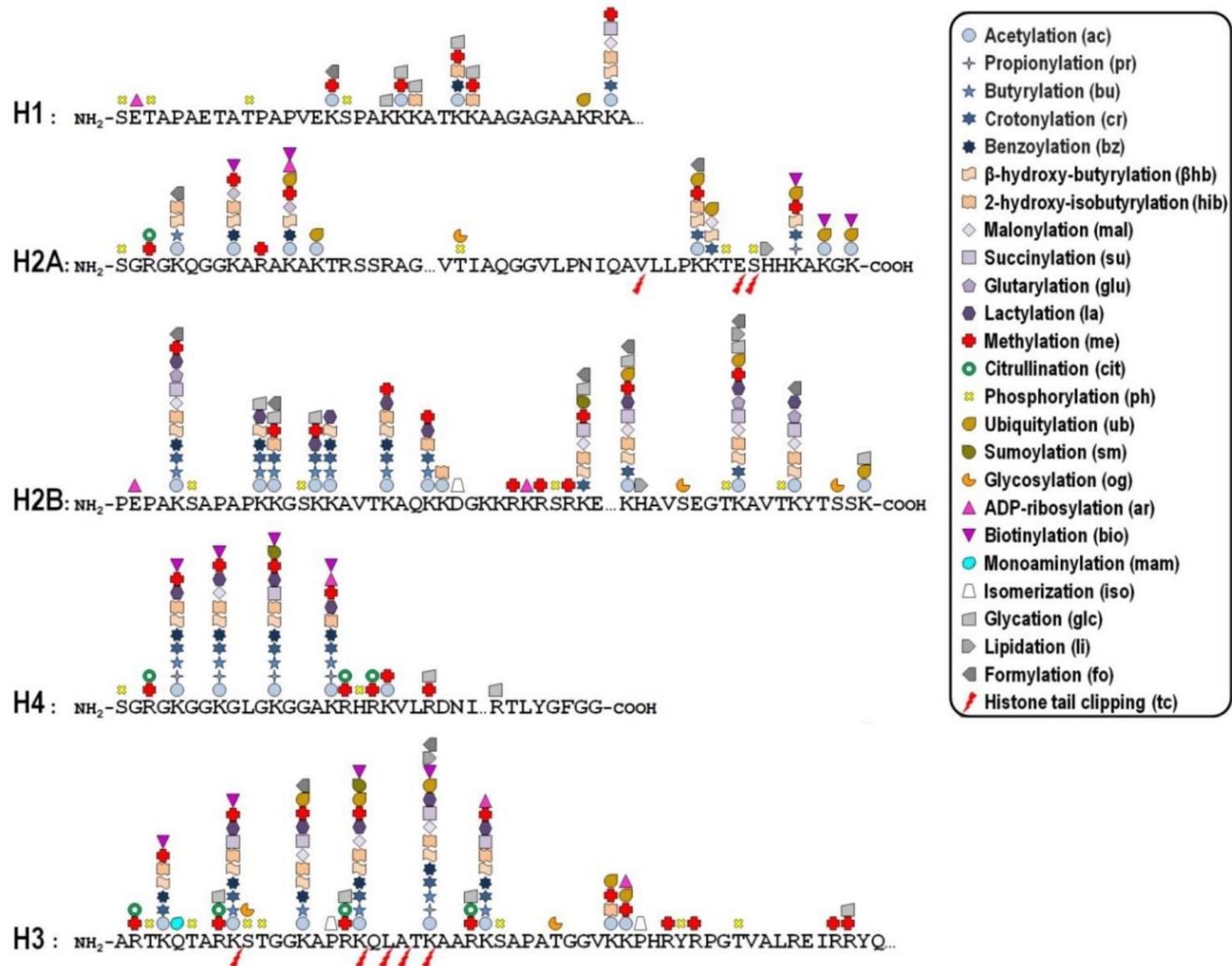
Post-transcriptional modifications of core histones



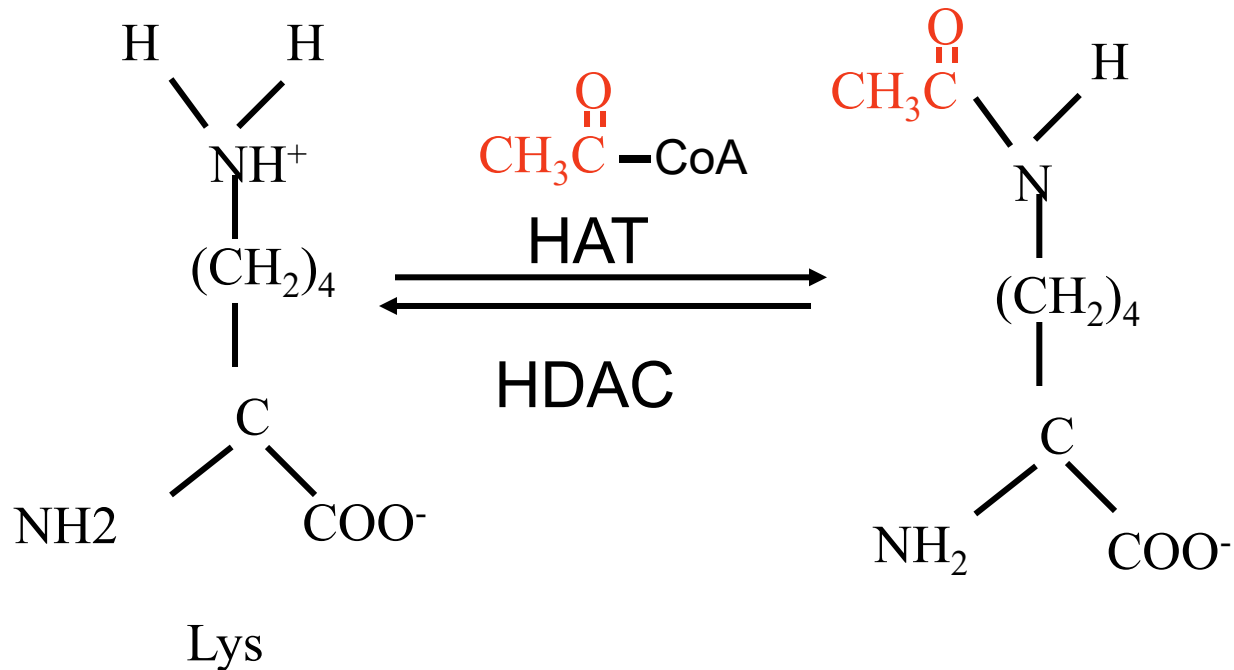
Adopted from Zhang and Reinberg, Genes & Dev (2001)

Not all histone modifications are epigenetically inherited.

Post-transcriptional modifications of core histones



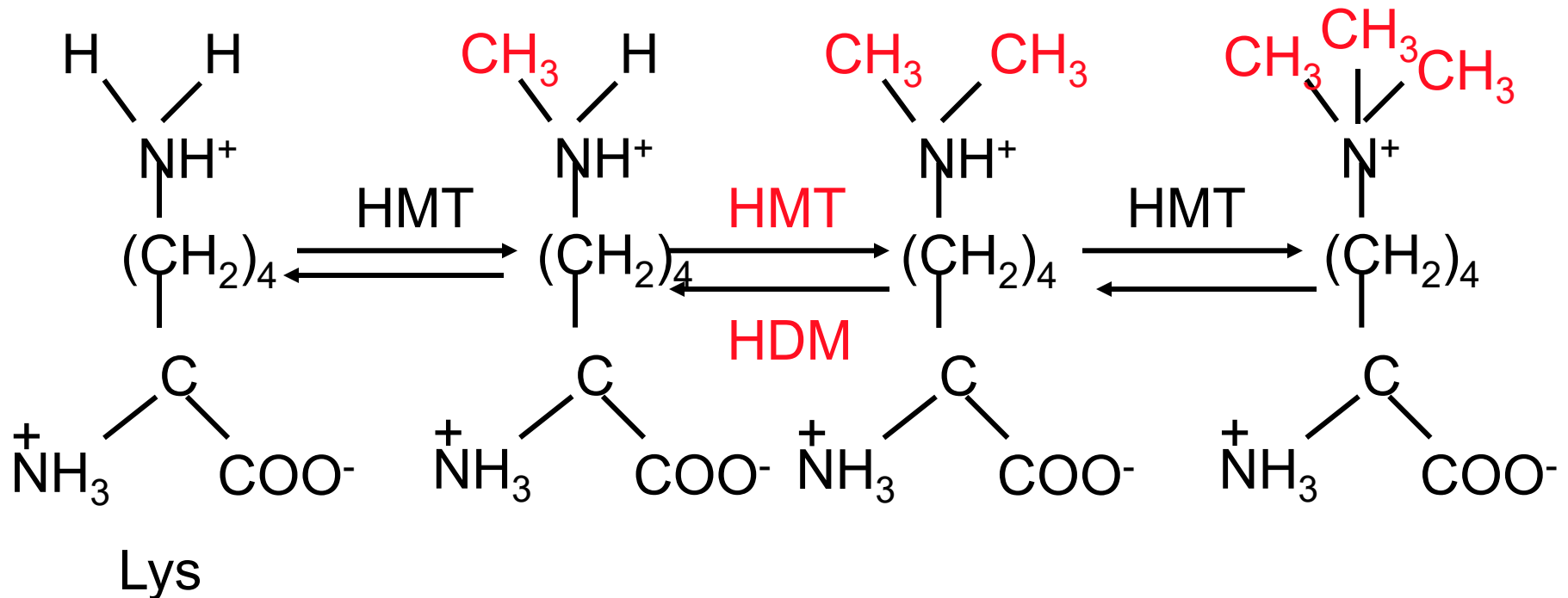
Acetylation of Lysine residue is reversible



HAT: histone acetyltransferase, more accurately; lysine acetyltransferase

HDAC: histone deacetylases

Lysine methylation is reversible

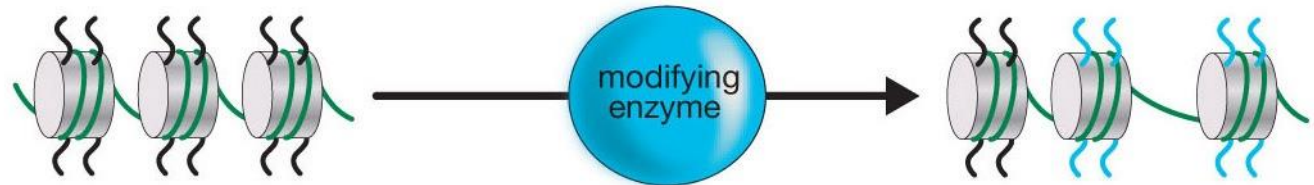


HMT: histone methyltransferase contains SET domain

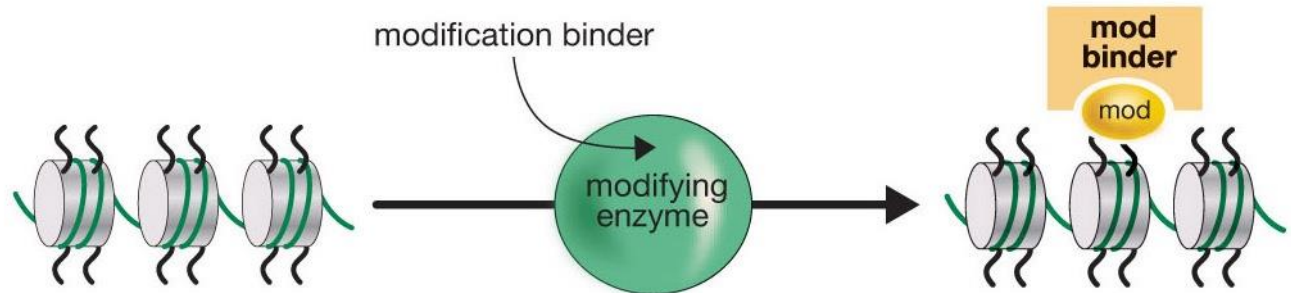
HDM: JmjC domains

What are the function of histone modifications?

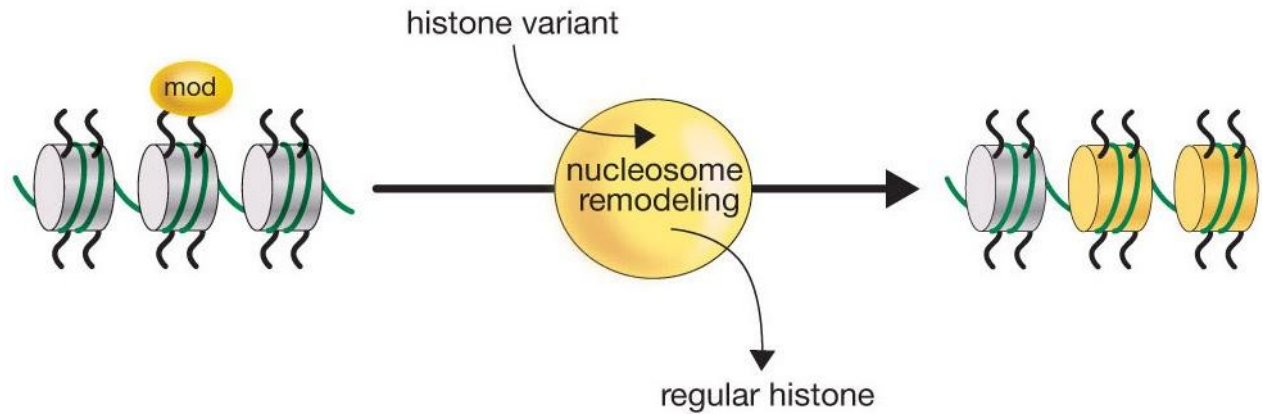
cis-effects



trans-effects

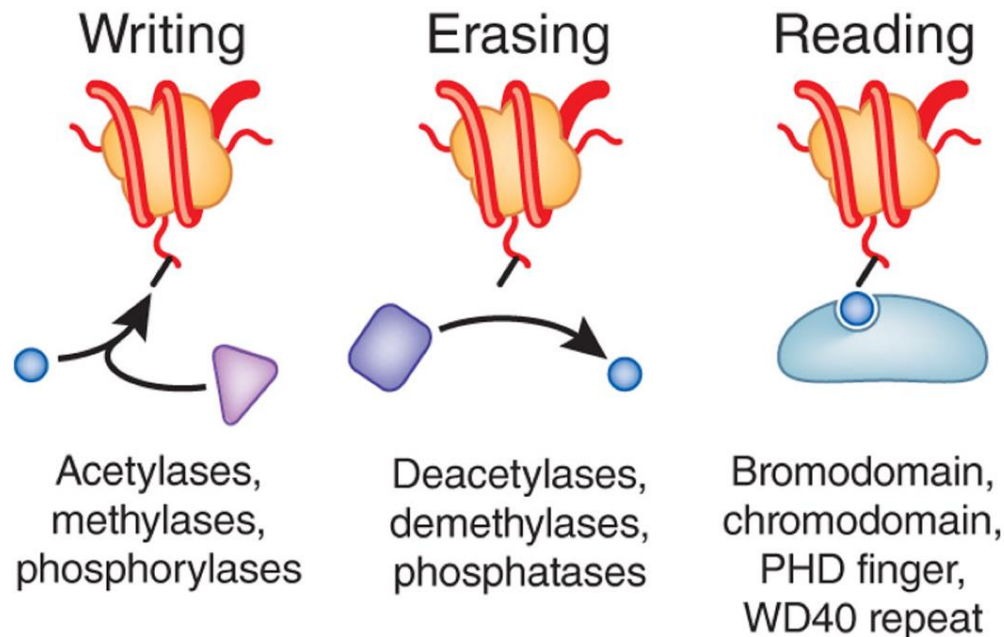


histone replacement

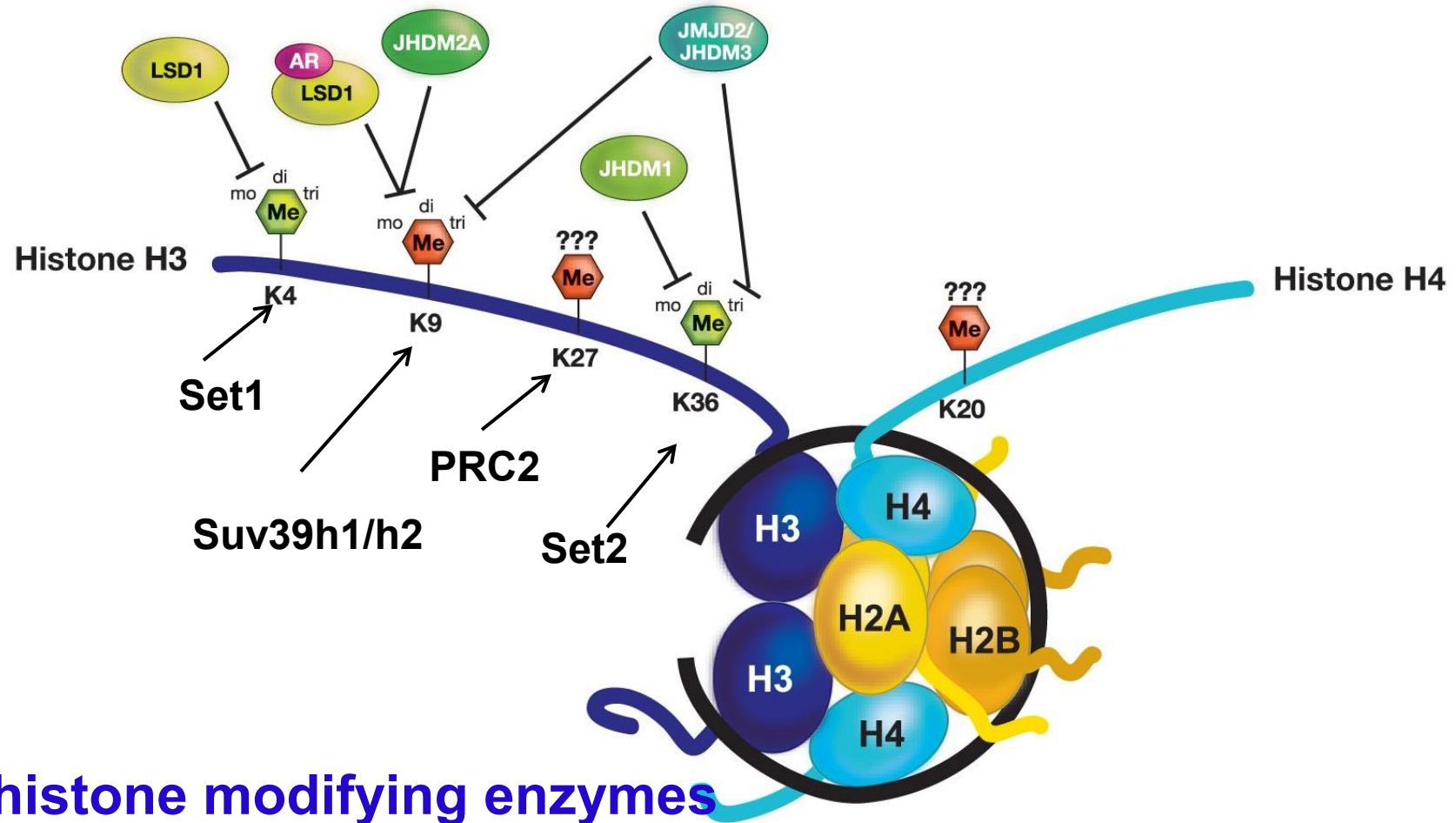


Three classes of proteins working on histone modifications

- **Writers:** enzymes that add a mark
- **Readers:** proteins that bind to and “interpret” the mark
- **Erasers:** enzymes that remove a mark

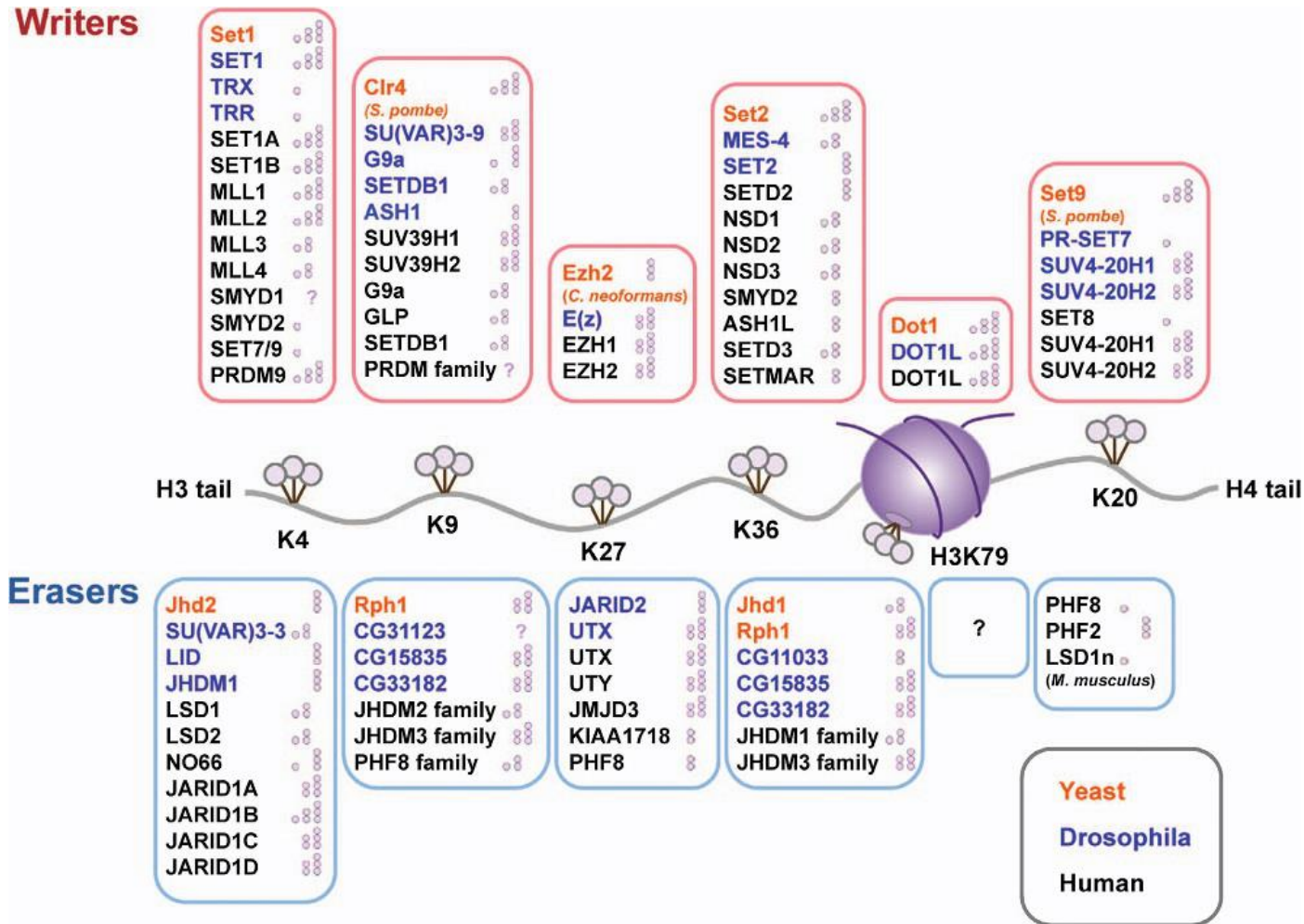


Examples of enzymes (writers) modifying the H3 tails



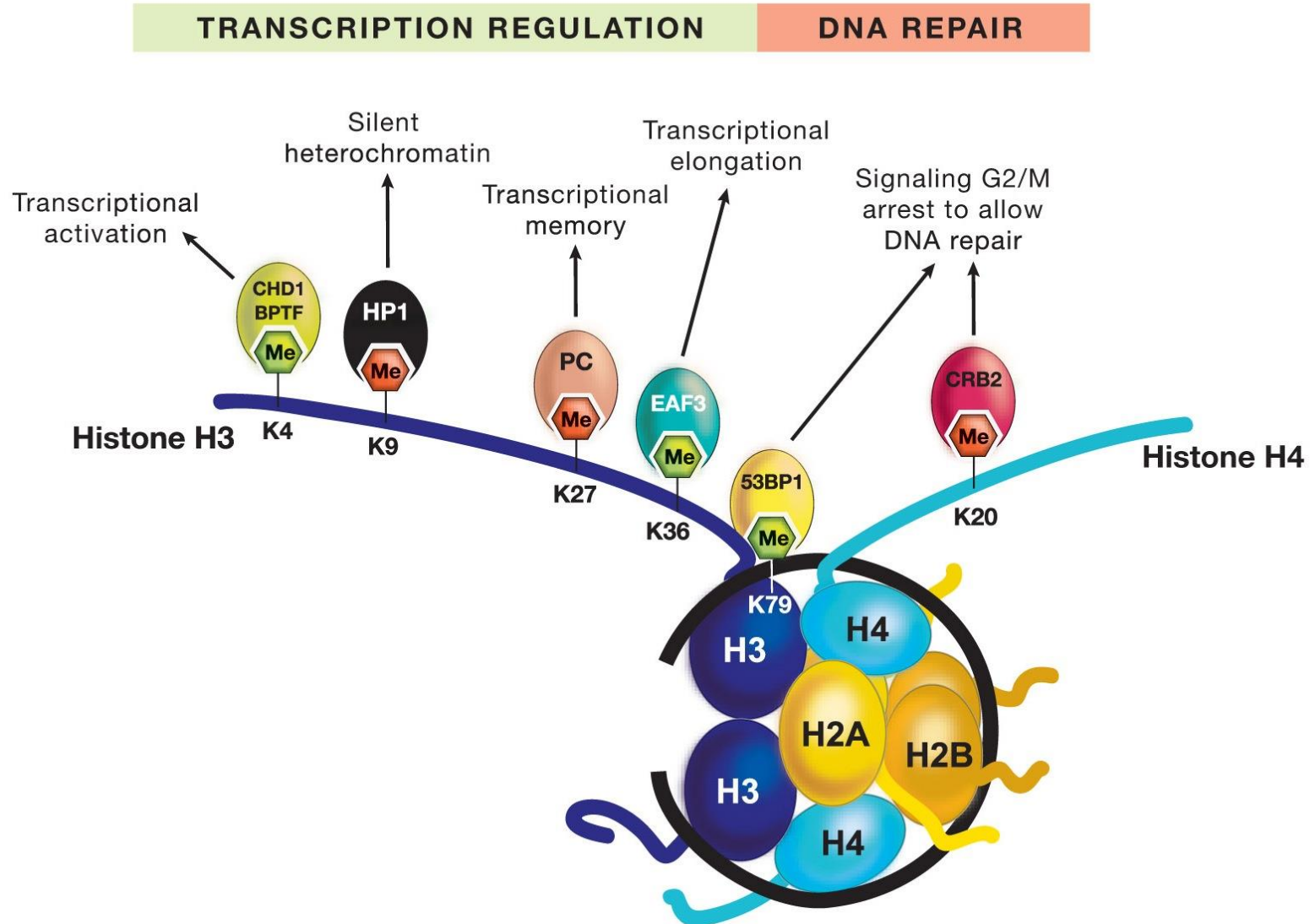
**Most histone modifying enzymes
have non-histone substrates**

Writers and Erasers for methylation of commonly studied lysine residues

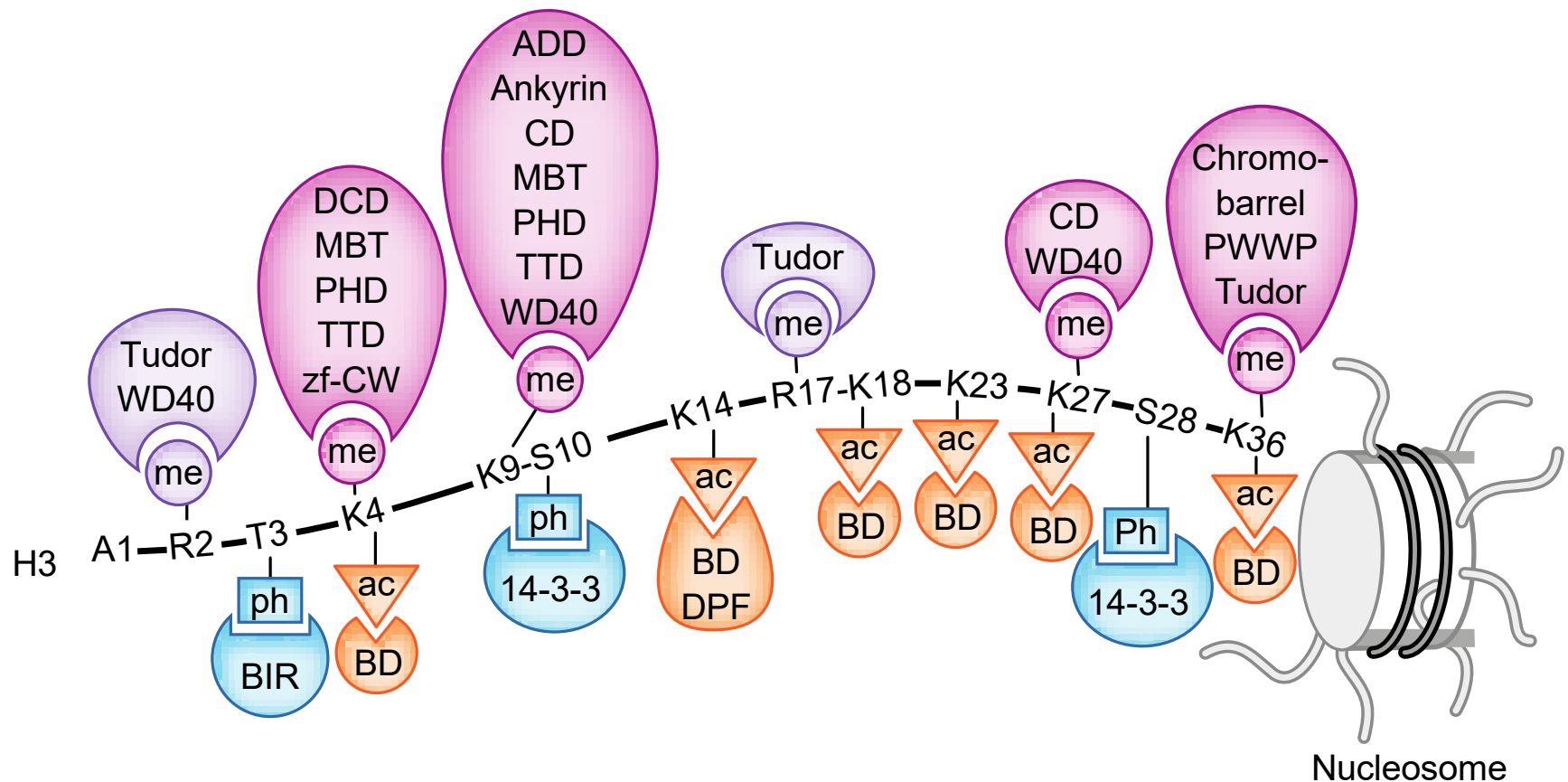


Most histone modifying enzymes have non-histone substrates

Proteins that read modifications on H3 and H4 tails

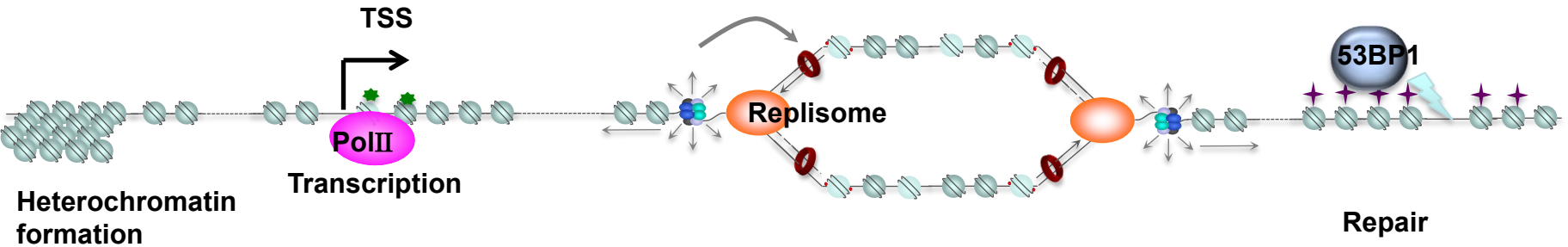


Proteins that read modifications on H3 and H4 tails



Enormous complexity and intricacy could be generated to regulate human epigenomes

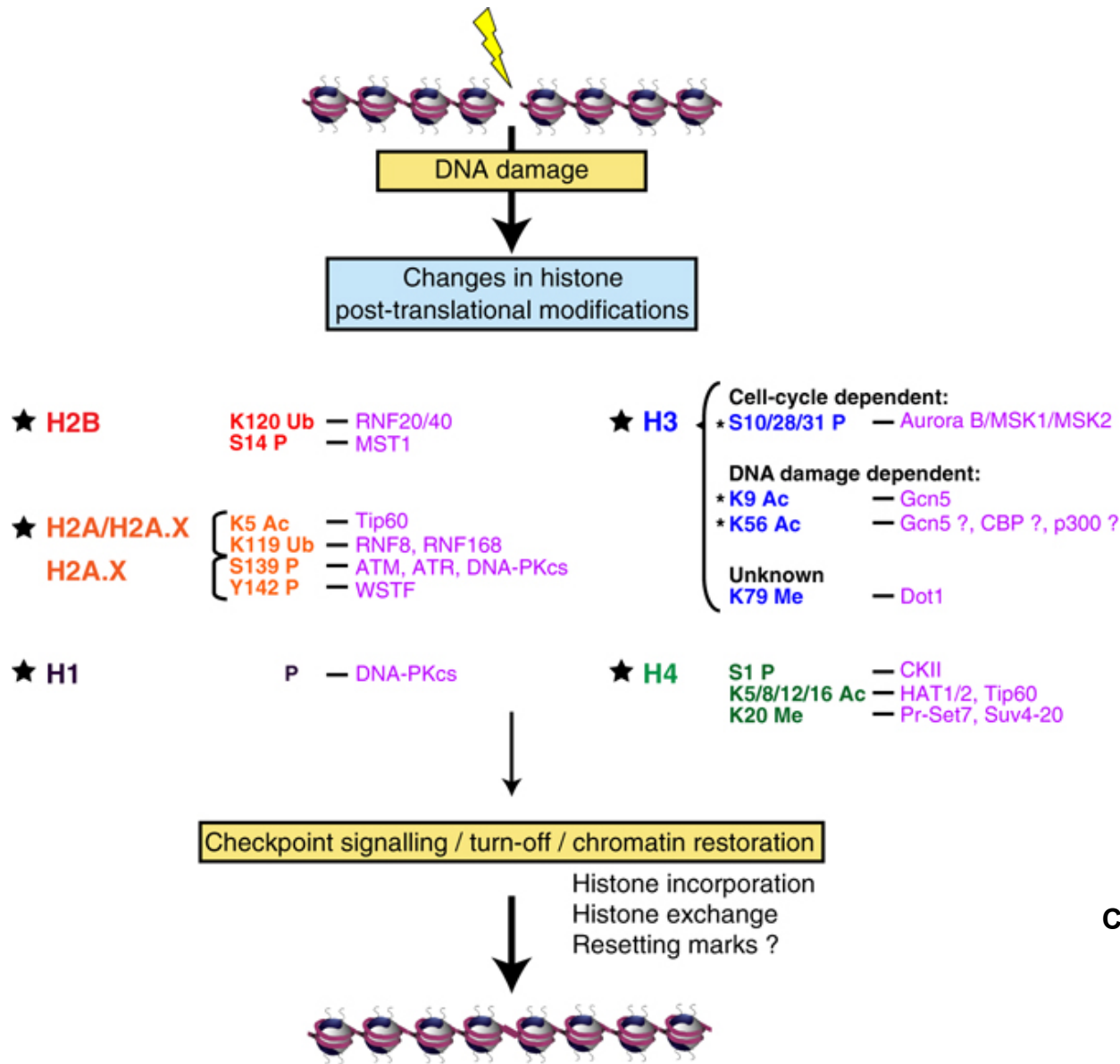
Histone modifications impact processes linking to DNA transactions



Active and inactive histone marks based on association with gene activity

- 1) Active marks: histone marks associated with active gene transcription (H3K4me1, H3K4me2, H3K4me3, H3K36me3, H3K9ac, H3K27ac, H4K16ac)**
- 2) Inactive/silent marks (H3K9me2 or H3K9me3, H3K27me3)**
- 3) Bivalent chromatin domains (H3K4me3 and H3K27me3) at developmentally regulated genes**

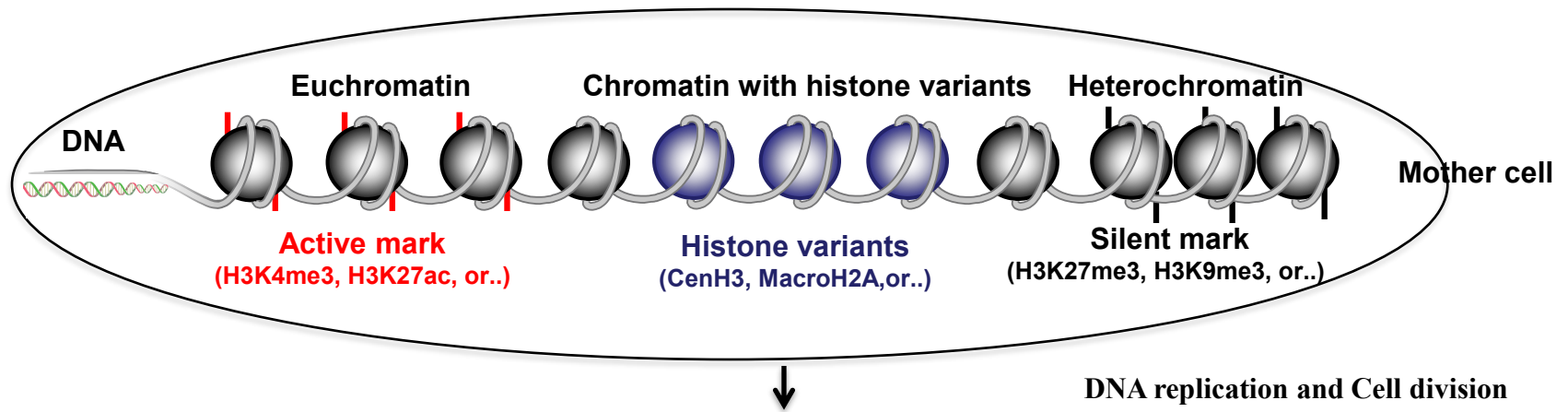
Histone modifications during DNA damage response



Epigenetic inheritance

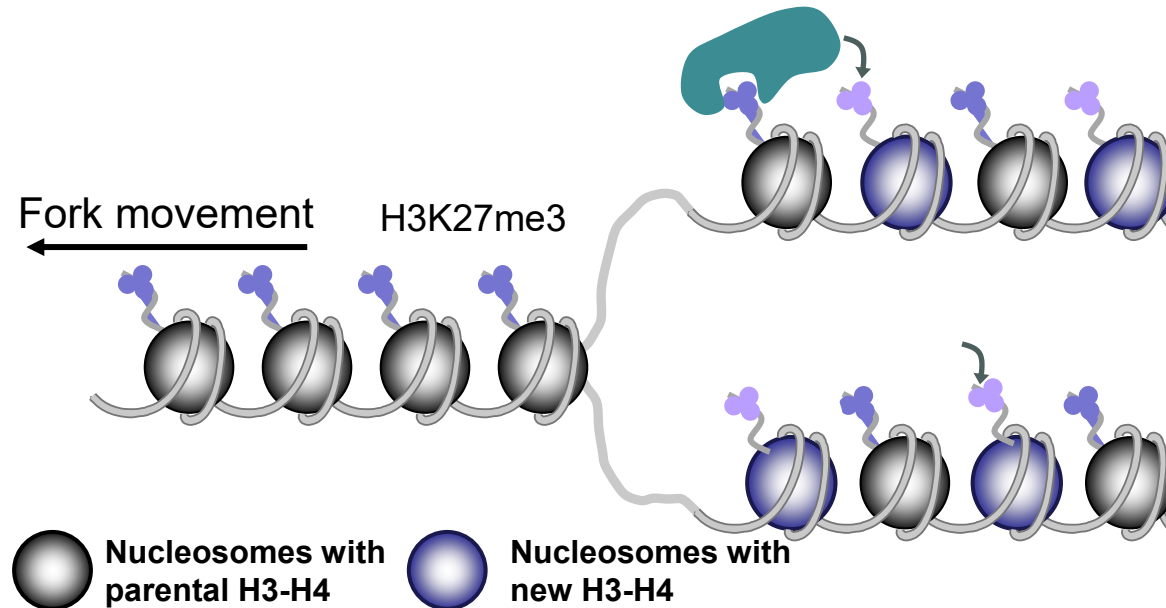
Epigenome duplication is also a daunting task

How are epigenetic states inherited during mitotic cell divisions



- Histone and histone variants are assembled at the same places
- Histone modification patterns at different chromatin domains are transmitted into daughter cells
- Readers for histone marks are recruited to the same places
- (Many others...)

DNA replication-coupled nucleosome assembly and the read and write mechanism for the restoration of histone marks



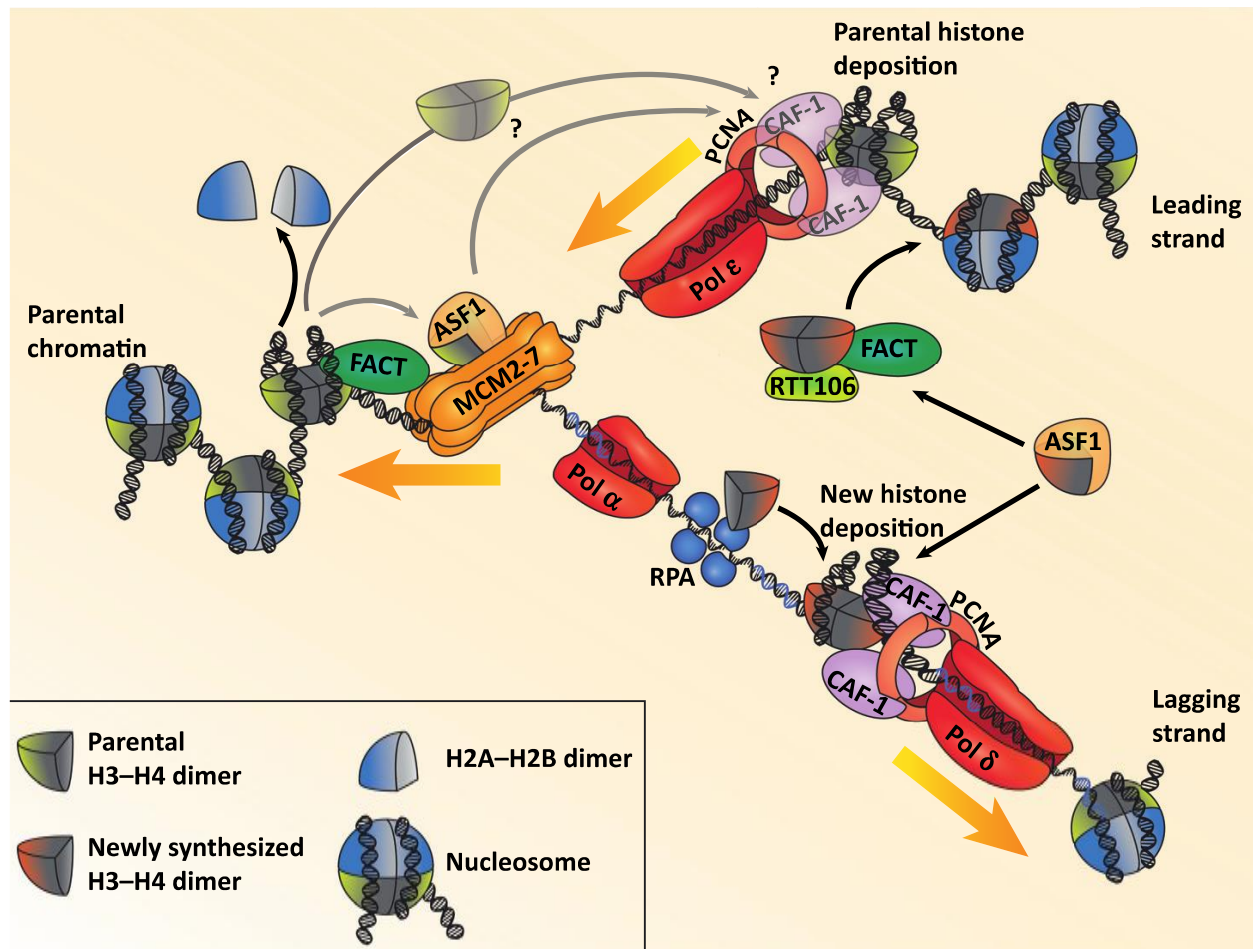
How are nucleosomes formed using newly synthesized H3-H4?

How are nucleosomes formed using parental H3-H4?

Du et al, Sci China Life Sci, 2022

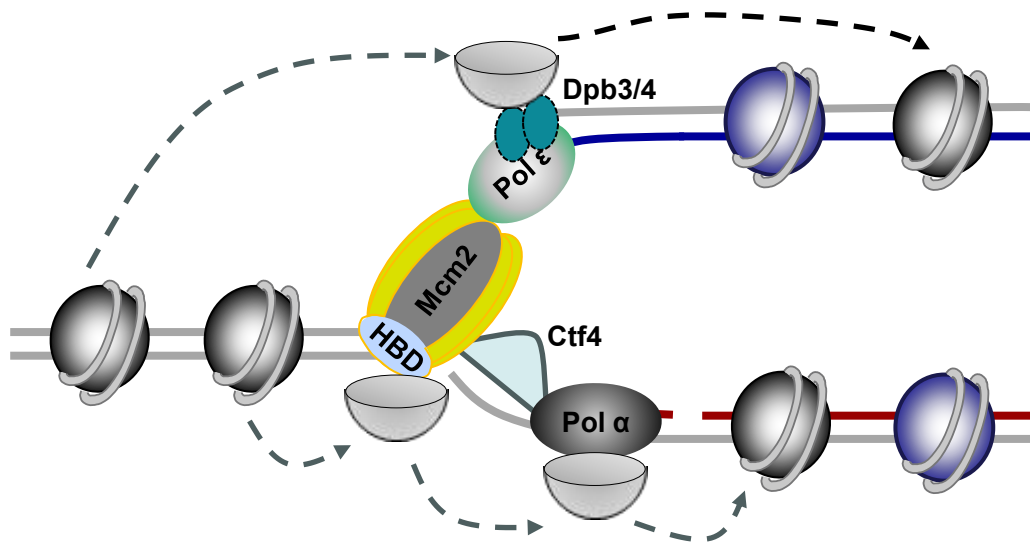
Escobar al, Nature Review Genetics, 2021

Many factors regulate the deposition of new H3-H4



Parental H3-H4 tetramers are transferred to replicating DNA strands via distinct mechanisms

Dpb3/4=POLE3/POLE4 in mammals



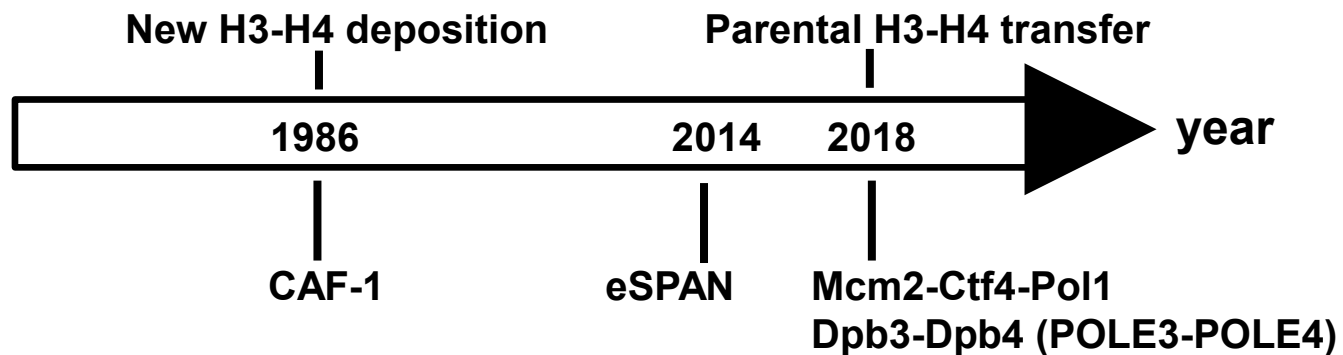
- Parental H3-H4 are transferred almost equally to leading and lagging strand of the DNA replication fork.
- Dpb3-Dpb4, two subunits of leading strand DNA polymerase, facilitates the transfer of parental H3-H4 tetramers to leading strand.
- The Mcm2-Ctf4-Pol α facilitates the transfer of parental H3-H4 to lagging strand.
- These pathways are conserved from yeast to mammalian cells.

Yu et al, Science 2018

Gan et al, Molecular Cell, 2018

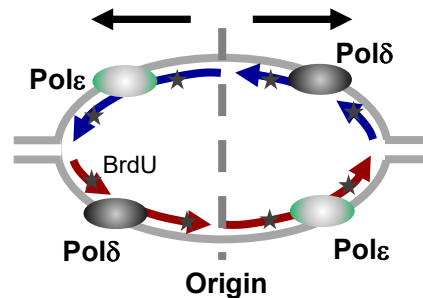
Petryk et al (Groth), Science, 2018

A simplified history of nucleosome assembly

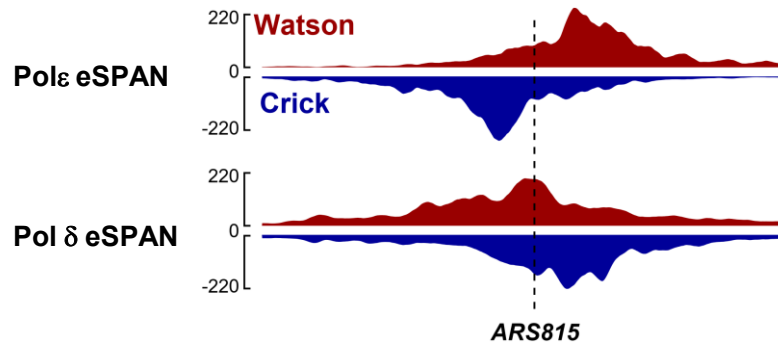


eSPAN can measure the relative amount of proteins at leading or lagging strands of DNA replication forks

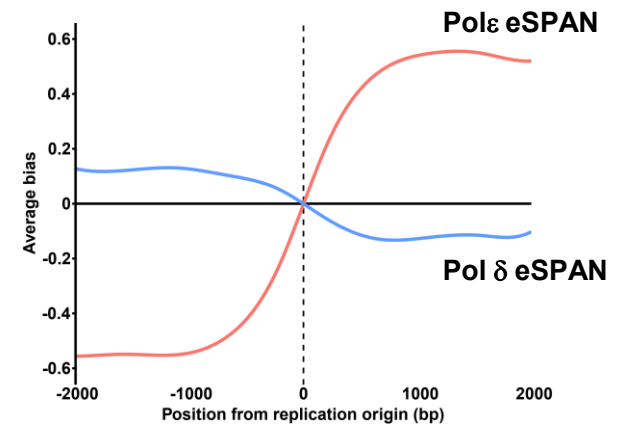
enrichment and
Sequencing of Protein
Associated Nascent DNA



Pursell et al (Kunkel), Science 2007
McElhinny et al (Kunkel), Mol Cell, 2008



$$\text{Bias} = (W-C)/(W+C)$$

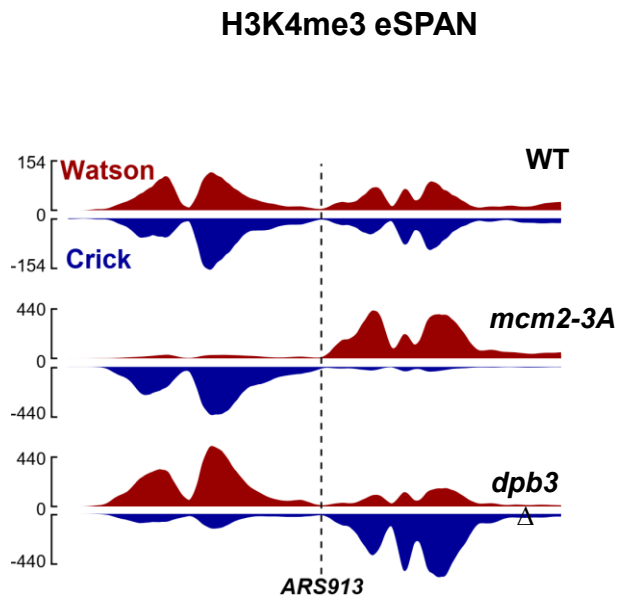


Bias pattern: leading vs lagging bias

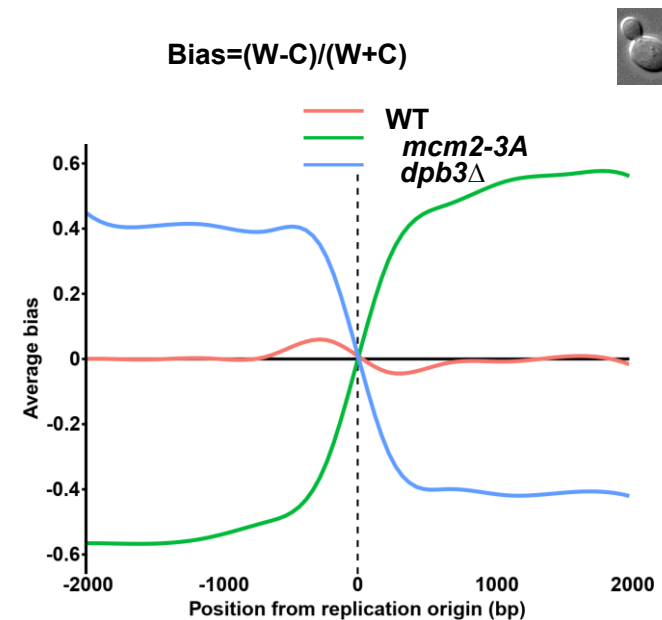
Bias ratio: relative amount of a protein at leading/lagging

Yu et al Molecular Cell 2014, PMCID: PMC436266

H3K4me3 eSPAN in *mcm2-3A* and *dpb3* Δ mutant cells exhibit a strong leading and lagging strand bias, respectively

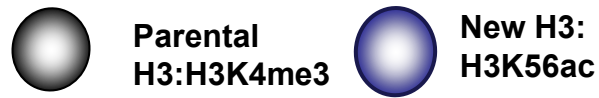


Dpb3 is a subunit of Pol ϵ .

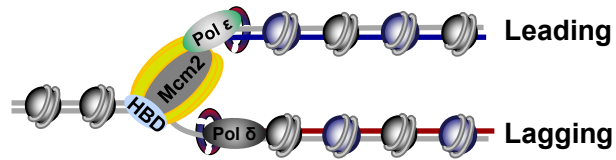


Yu et al, Science 2018, PMCID: PMC6597248

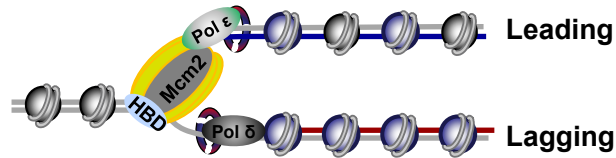
Gan et al, Molecular Cell, 2018, PMCID: PMC6193272



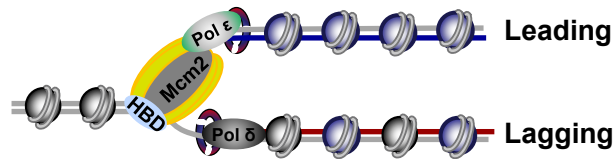
WT



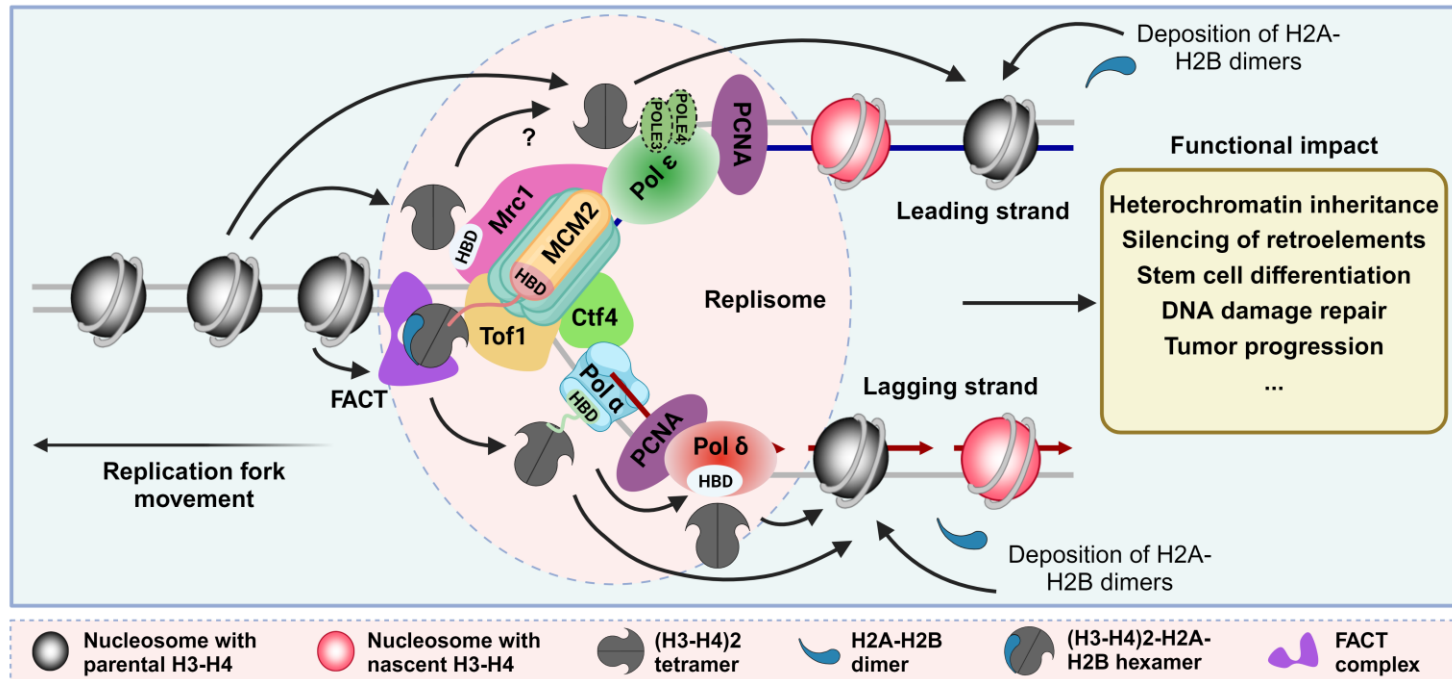
mcm2-3A



dpb3Δ



Multiple replisome components likely work in relay fashion to transfer parental histones to replicating DNA strands



Petryk et al, Science, 2018
 Dolce et al, Genes Dev, 2022
 Wang et al, NAR 2023
 Wen et al, Nature genetics, 2023
 Weger et al, Nature Genetics, 2023
 Yu et al, Cell, 2024
 Toda et al, Mol Cell, 2024
 Li et al, Nature, 2024
 Charlton et al, Cell, 2024
 Tian et al, PNAS, 2024
 Shi et al, Science Advances, 2024
 Karri et al, NAR, 2024

Yu et al, Science, 2018
 Gan et al, Molecular Cell, 2018
 Li et al, Science Advances, 2019
 Serra-Cardona et al, Science Advances, 2022
 Xu et al, eLife 2022
 Xu et al, Nature Communications, 2022
 Li et al, Nature 2023
 Fang et al, Genes Dev, 2024
 Serra-Cardona et al, Science Advances, 2024

Replisome components are responsible for faithful duplication of both genetic and epigenetic information

The nucleosome core particle remembers its position through DNA replication and RNA transcription

Gavin Schlissel^a and Jasper Rine^{a,1}

^aDepartment of Molecular and Cell Biology, University of California, Berkeley, CA 94720

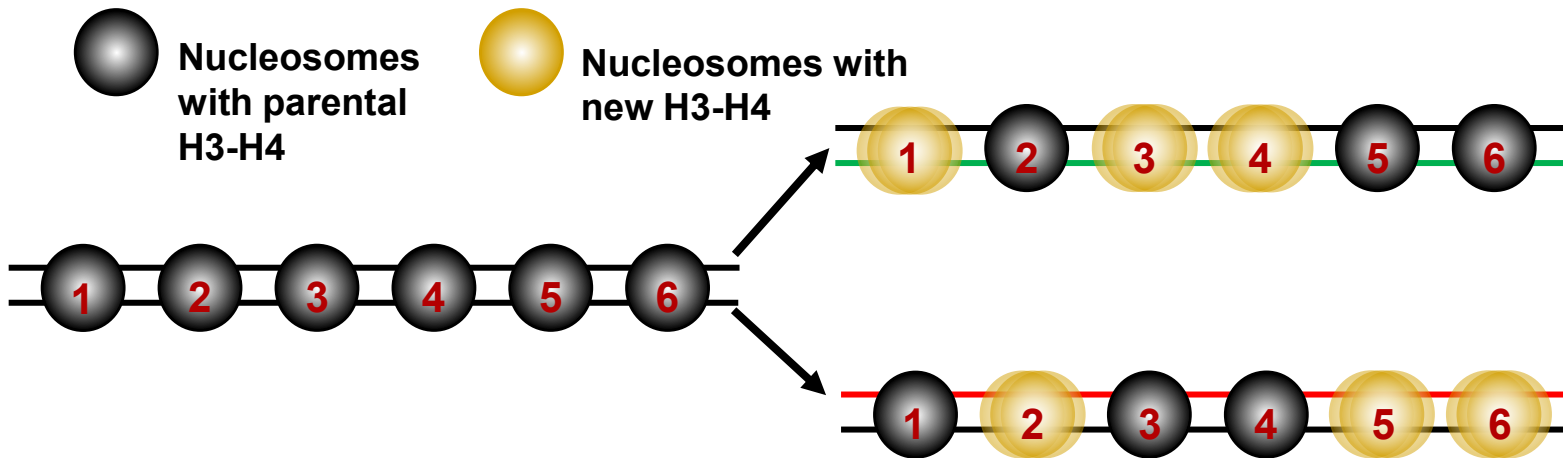
Contributed by Jasper Rine, August 9, 2019 (sent for review July 12, 2019; reviewed by Steven Henikoff and Fred M. Winston)

Article

Cell

Active and Repressed Chromatin Domains Exhibit Distinct Nucleosome Segregation during DNA Replication

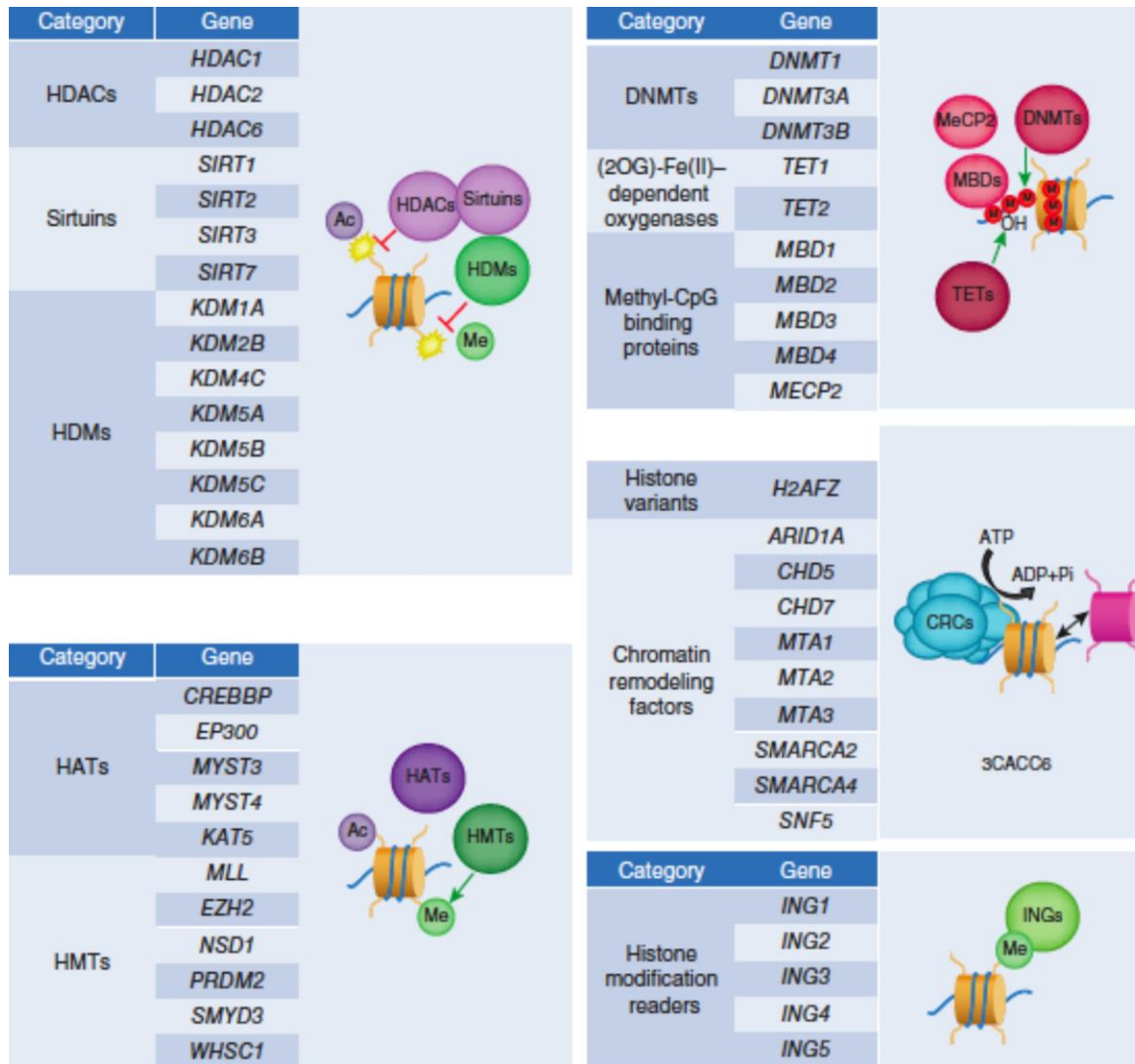
Thelma M. Escobar,^{1,2} Ozgur Oksuz,^{1,2,3} Ricardo Saldaña-Meyer,^{1,2} Nicolas Descostes,^{1,2,4} Roberto Bonasio,^{1,2,5} and Danny Reinberg^{1,2,6,*}



Epigenetics and Cancer

- **Cancer is a disease caused by both genetic and epigenetic changes**

Chromatin regulators are altered in a variety of tumors



Article

Transient loss of Polycomb components induces an epigenetic cancer fate

<https://doi.org/10.1038/s41586-024-07328-w>

Received: 20 January 2023

Accepted: 15 March 2024

Published online: 24 April 2024

Open access



Check for updates

V. Parreno^{1,9}, V. Loubiere^{1,2,9}, B. Schuettengruber¹, L. Fritsch¹, C. C. Rawal³, M. Erokhin⁴, B. Györfy^{5,6}, D. Normanno¹, M. Di Stefano¹, J. Moreaux^{1,7,8}, N. L. Butova³, I. Chiolo³, D. Chetverina⁴, A.-M. Martinez¹✉ & G. Cavalli¹✉

Although cancer initiation and progression are generally associated with the accumulation of somatic mutations^{1,2}, substantial epigenomic alterations underlie many aspects of tumorigenesis and cancer susceptibility^{3–6}, suggesting that genetic mechanisms might not be the only drivers of malignant transformation⁷. However, whether purely non-genetic mechanisms are sufficient to initiate tumorigenesis

Histone modifications and cancer

Many chromatin regulators are frequently mutated in cancer cells

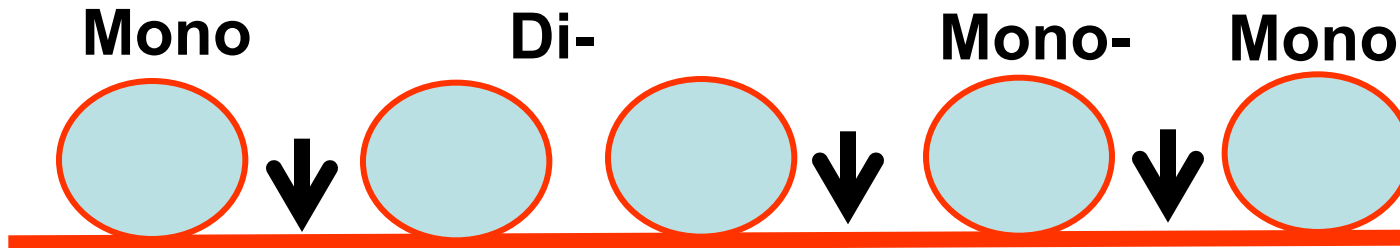
Mutations at gene regulatory elements (enhancers, promoters) are detected in a variety of cancers

A global change in histone modifications has been detected in cancer cells

Expression of histone modifying enzymes is altered in cancer cells

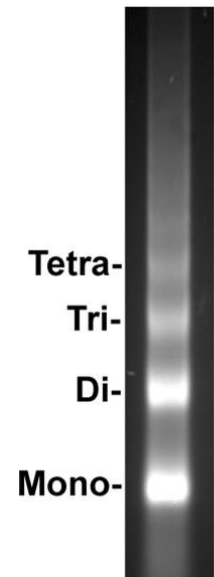
**How to probe chromatin and histone
modifications?**

Analysis of nucleosome positioning by Mnas-seq



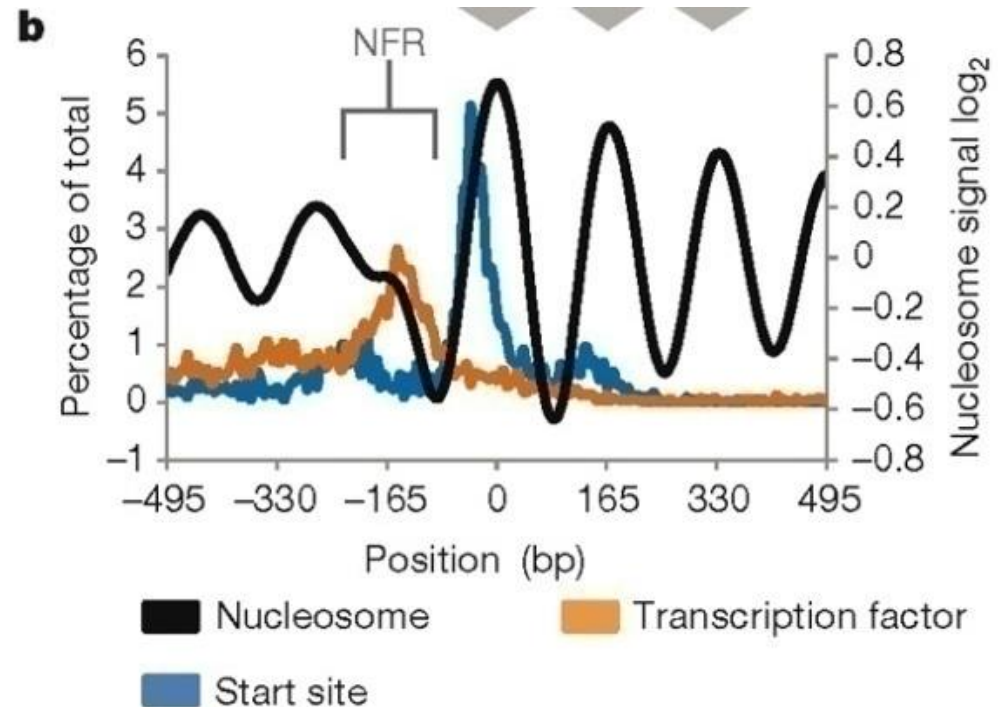
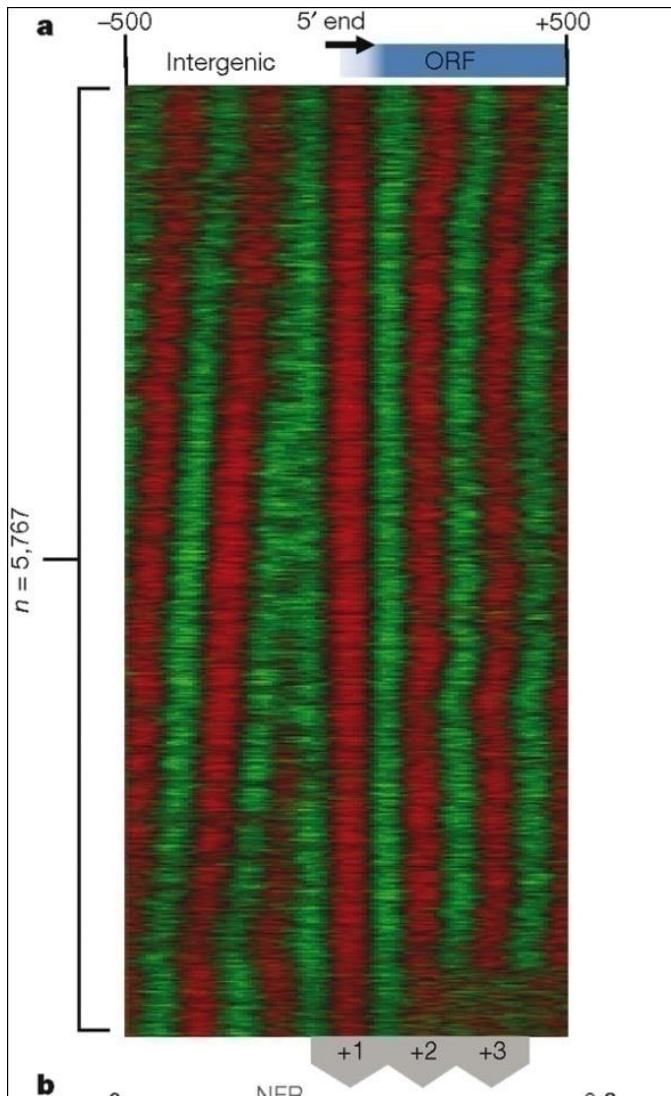
Basic repeating structure can be probed
(protect and seq method)

- Digestion enzyme cuts accessible
regions of DNA



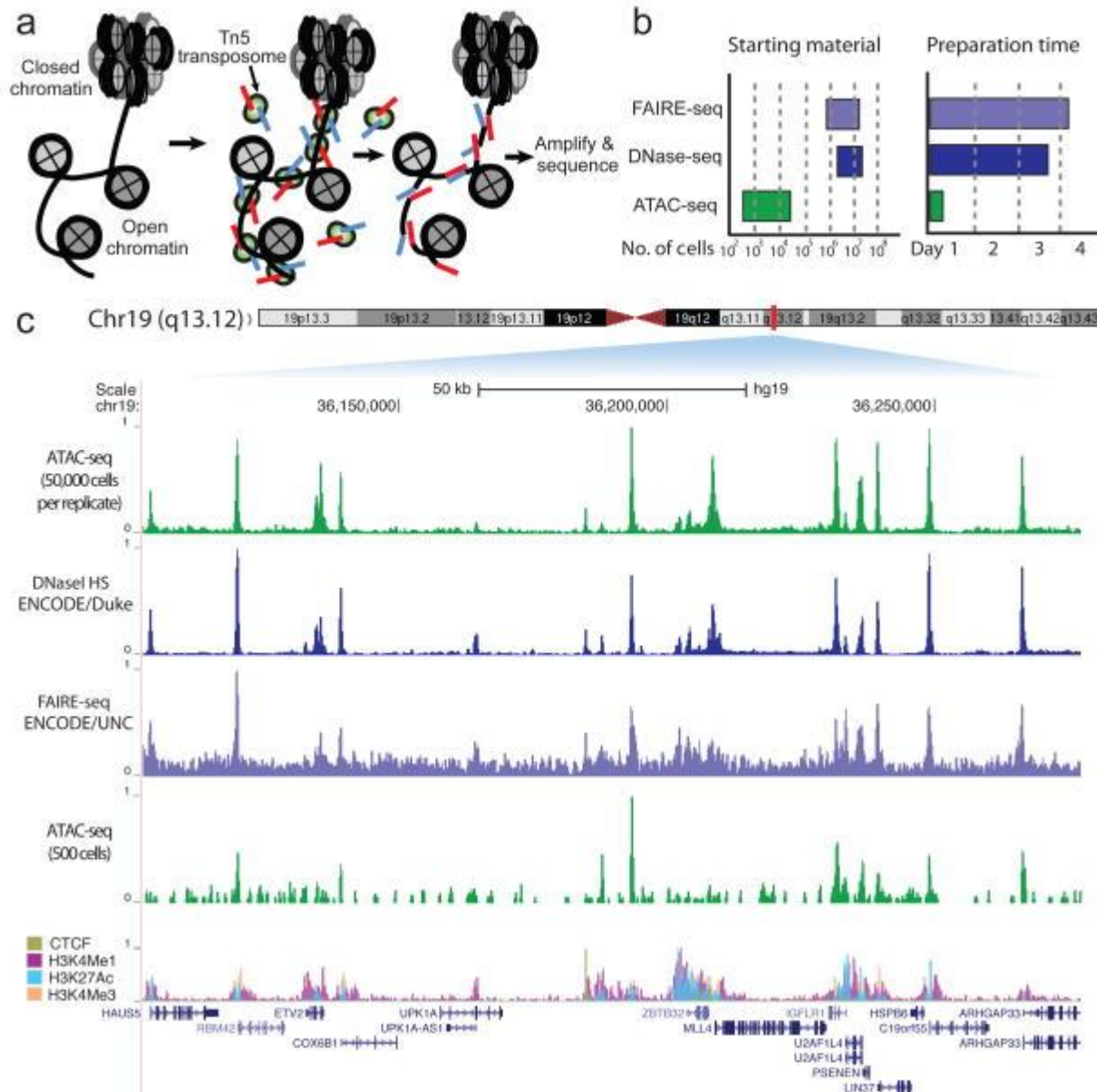
Nucleosome positioning

Gene regulatory elements such as promoters and enhancers are at “nucleosome free”



Whitehouse et al., 2007 Nature 450, 1031-1035

Analysis of nucleosome free region/open chromatin by ATAC-seq



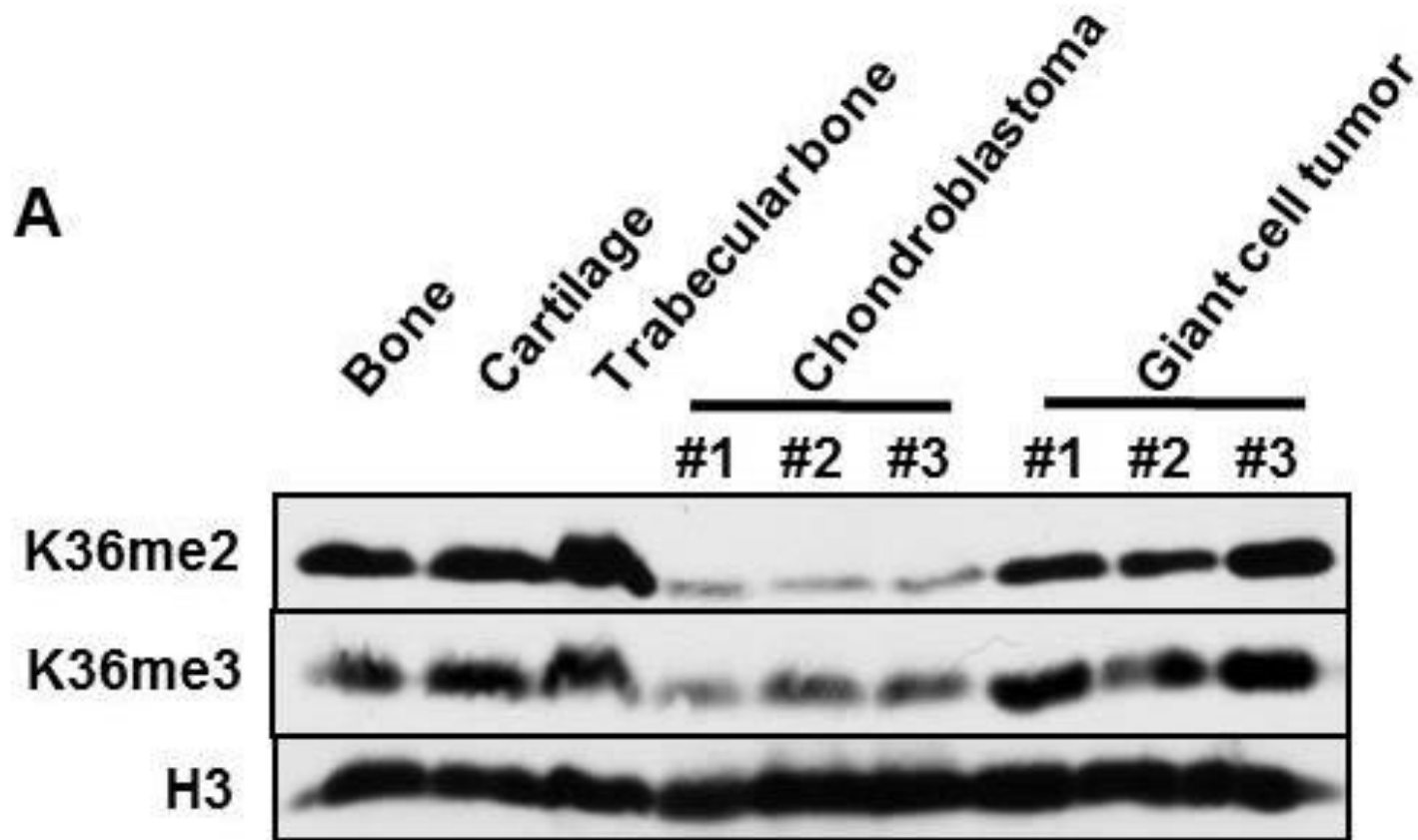
Analysis of histone modification by Western blot and ChIP-seq

Chondroblastoma

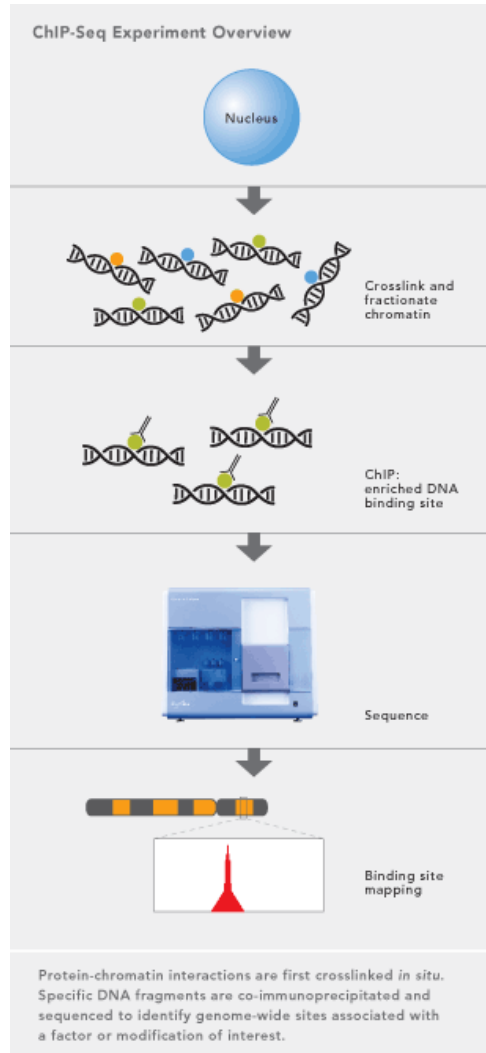


- arises from the epiphysis of the long bones
- Characterized by high cellularity and undifferentiated tissues
- over 90% cases contain H3.3K36M mutation
- The molecular mechanism of tumorigenesis is unknown

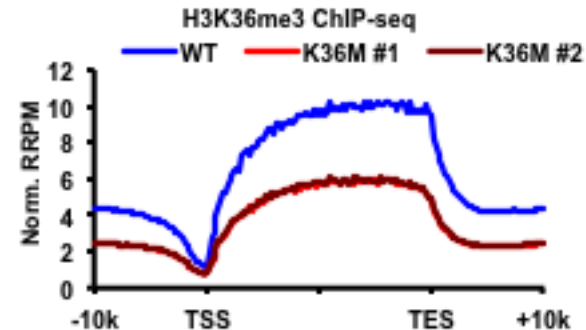
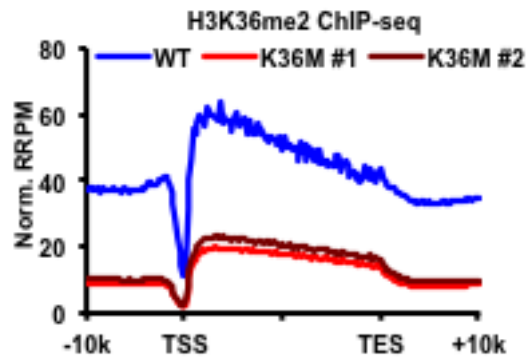
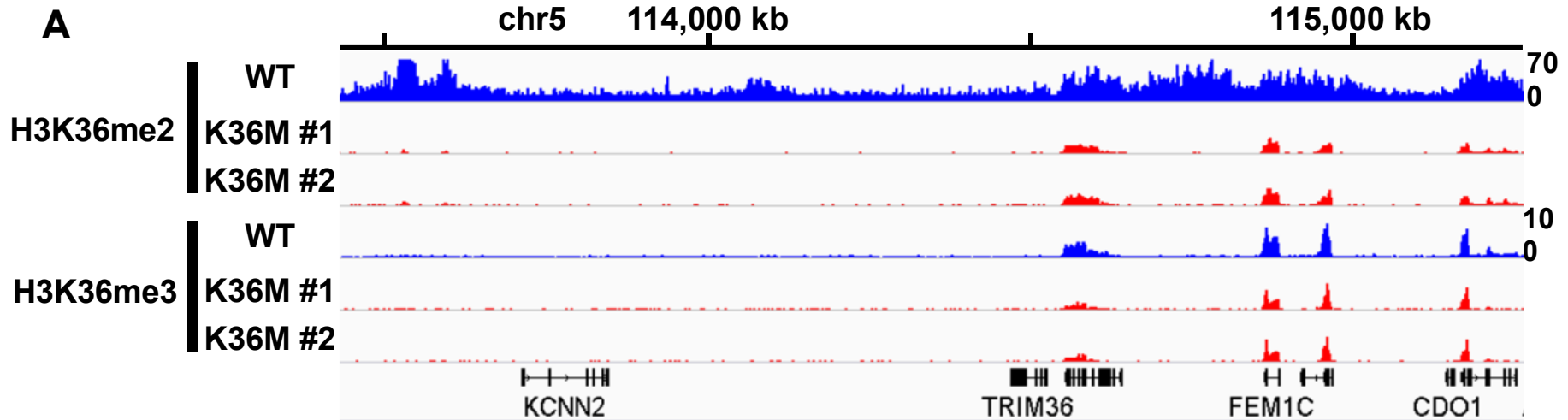
H3K36me2/me3 levels are low in chondroblastoma samples compared to normal tissue samples



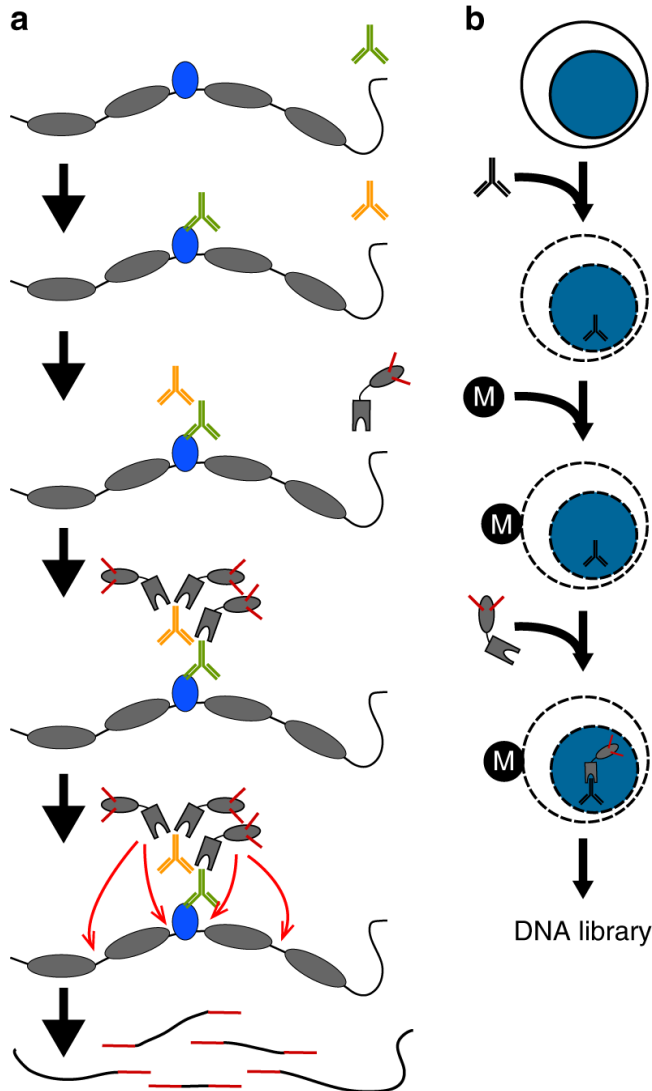
Map histone modifications to a DNA specific sequence by ChIP-seq



Effect of H3.3K36M mutation on H3K36me2 and H3K36me3 on chromatin in cell lines



(CUT&tag): a new method for epigenomic profiling



Advantages compared to ChIP-seq

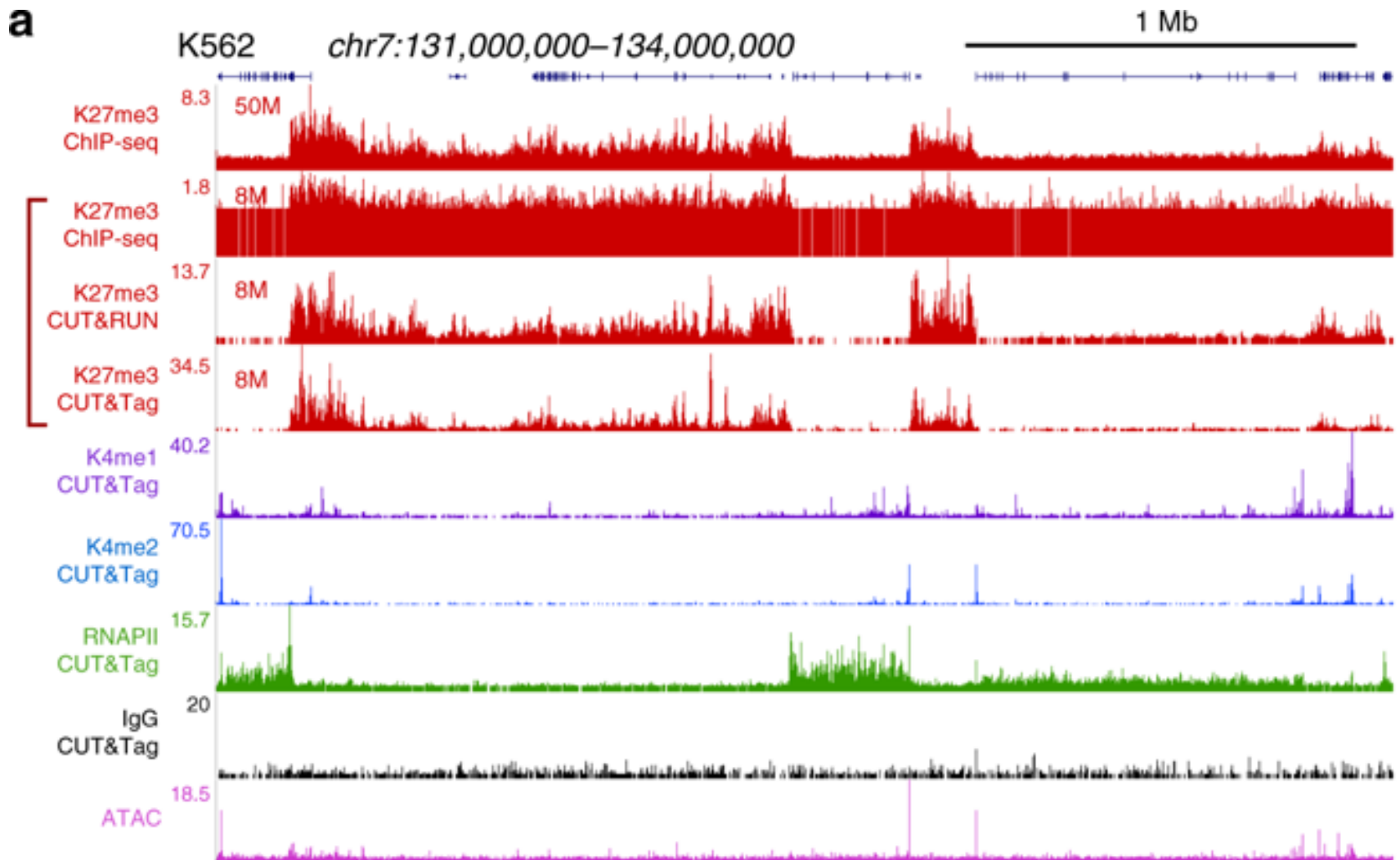
- **Fast and efficient**
- **Low background and therefore less sequence reads**
- **A dramatic reduction of sequencing cost**
- **Low cell number**

Potential issues:

- **complication of ATAC-seq signals**

Kaya-Okur et al Nature Communications 2019
Carter et al Nature Communications 2019

Cleavage Under Targets and Tagmentation (CUT&tag) (ACT-seq): a new way for epigenomic profiling



Cleavage Under Targets and Release Using Nuclease (CUT&RUN): a method for epigenomic profiling

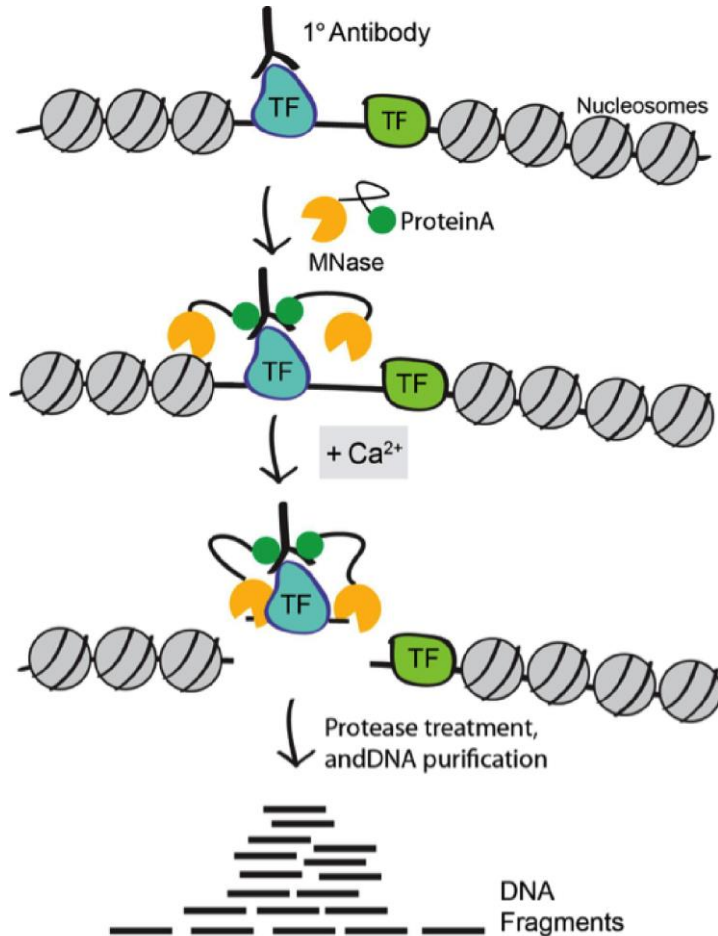


Figure 1. CUT&RUN schematic (see text for details).

Advantages compared to CUT&tag

- No potential ATAC-seq signals
- Detect transcription factors

Disadvantages compared to CUT&tag

- Need a library preparation kits
- Maybe better more cells

Take home messages

- **Epigenetic phenomenon occurs in our daily life.**
- **Mechanisms of epigenetic regulations are complex and evolving.**
- **Basic principles of epigenetic regulations have been defined.**
- **Tools to probe histone modifications and chromatin structures are evolving at single cell and single molecule levels.**